

Research Article

**MORPHOLOGY AND PHYLOGENY OF *RETIBOLETUS KAUFFMANII*
(BOLETACEAE): A NEW RECORD OF WILD MUSHROOM
FROM INDIAN HIMALAYA**

D. Chakraborty¹, *Kanad Das¹, A. Parihar¹, S. Adhikari², S. K. Mukherjee², J. Karmakar³ and T. K. Bandyopadhyay³

¹*Cryptogamic Unit, Botanical Survey of India, P.O. – Botanic Garden, Howrah 711103, India*

²*Department of Botany, University of Kalyani, Kalyani 741235, West Bengal, India*

³*Department of Molecular Biology & Biotechnology, University of Kalyani, Kalyani 741235, West Bengal, India*

**Author for Correspondence*

ABSTRACT

Retiboletus kauffmanii is reported for the first time from India. Details of macro- and micro-morphology along with nrLSU-based phylogenetical analysis are presented for the Indian collection. Similar looking Asian or extra limital species are compared.

Keywords: *Boletales, Macrofungi, Phylogenetic Study, Sikkim, Taxonomy*

INTRODUCTION

Continuous development in molecular phylogeny corresponds with the revolutionary changes on the systematic studies of boletes in recent years (Nuhn *et al.*, 2013; Wu *et al.*, 2014, 2016). The family Boletaceae (one of the 17 families in Boletales) alone has a worldwide representation of 66 generic clades including nearly 1050 species (Li *et al.*, 2016; Wu *et al.*, 2016). The genus *Retiboletus* Binder & Bresinsky (2002) represented by one of these 66 generic clades is mainly characterised morphologically by: dry to subtomentose, black, dark gray, mustard yellow to olive brown colored pileus; pallid, grayish to yellow tubes and pores; centrally placed stipe with reticulation; pallid, yellow or vividly yellow context that changes slightly or to orange brown on bruising; smooth basidiospores; presence of cystidia and absence of clamp connections. Recent molecular phylogeny of the family Boletaceae placed them under subfamily *Leccinoideae* i.e. clade 15 (Wu *et al.*, 2014). Combination of molecular phylogenetic and morphological studies also clearly shows the existence of 13 distinct species from all over the world (Zeng *et al.*, 2016).

Sikkim (one of the Himalayan states in India) which comes under one of the 18 biodiversity hot spots of the world was seriously under explored in terms of boletes until recent past (since 2008) when Botanical Survey of India undertook the extensive and intensive macrofungal explorations of fleshy wild mushrooms. During a routine foray to South and East district of Sikkim in 2016 two (KD & DC) of us collected a number of Boletes. Thorough morphological examination followed by LSU-based phylogenetic analysis of these mushrooms revealed one of them as *Retiboletus kauffmanii*, a recently established species from China. We report this species for the first time from India.

MATERIALS AND METHODS

Morphology

Macromorphological features were recorded from the fresh basidiomata in the field, including macro chemical (with KOH, FeSO₄ and Guaiacol) colour-reactions, prior to drying with a portable dryer. Photographs of fresh basidiomata and microphotographs were taken with Cannon SX 220 HS and Nikon-DS-R1 (dedicated to Nikon Eclipse Ni compound microscope) cameras. Colour codes and terms mostly follow Kornerup & Wanscher (1978). Micro morphological characters were recorded with the help of compound microscope (Nikon Eclipse Ni) from free hand sections of dry samples mounted in 5 % KOH, or stained in a mixture of 5 % KOH and phloxin and mounted in 30 % glycerol. Drawings were made with a drawing tube (attached to Nikon Eclipse Ni microscope) at 1000×. Basidium length excludes

Research Article

length of sterigmata. Spore-measurements were recorded in profile view from twenty basidiospores. Spore-size measurements and length/width ratios (Q) are given as: minimum–mean–maximum. Herbarium codes follow Thiers (continuously updated).

DNA Extraction, Polymerase Chain Reaction (PCR) and Sequencing

Genomic DNA was extracted from 100 mg of dried basidiome using the InstaGene™ Matrix Genomic DNA isolation kit (Biorad, USA) following the manufacturer's instructions. The Nuclear LSU region was amplified with primer pairs LR0R and LR5 (Vilgalys & Hester 1990; <http://www.biology.duke.edu/fungi/mycolab/primers.htm>). PCR amplification was performed on a thermal cycler (Eppendorf, Germany) programmed for 2 min at 94°C, followed by 40 cycles of 30 sec at 94°C, 30 sec at 50°C, 45 sec at 72°C and a final stage of 5 min at 72°C. The PCR product was purified using the QIAquick PCR Purification Kit (QIAGEN, Germany) and directly sequenced on the 3730xl DNA Analyzer (Applied Biosystems, USA) using the amplifying primers. The newly generated sequence was deposited at GenBank to procure the accession number (KY290586).

Phylogenetic Analysis

Phylogenetic analyses based on nrLSU sequence data were carried out to establish the phylogenetic placement of our isolated taxon. The reference sequences and outgroup taxa were selected from relevant literature (Zeng *et al.*, 2016; Desjardin *et al.*, 2009) and information retrieved from GenBank. All sequences were aligned with MAFFT v. 7 (Kato *et al.*, 2002). No manual editing was done within the alignment. Phylogenetic analysis was conducted with Maximum Likelihood (ML) method implemented in MEGA 6.0 (Tamura *et al.*, 2013). One-thousand bootstrap replicates were analysed to obtain nodal support values. *Spongiforma thailandica* and *S. squarepantsii* were chosen as out group taxa.

RESULTS AND DISCUSSION

Phylogeny

Our LSU-based phylogenetic analysis (Figure 1) with 34 LSU sequences (including the present species) resolved genus *Retiboletus* with full support. This analysis clearly supports the studies undertaken by Zeng *et al.*, (2016) indicating the existence of 13 species level subclades within the genus *Retiboletus*. Out of existing 13 subclades six (*R. sinensis*, *R. kauffmanii*, *R. zhangfeii*, *R. pseudogriseus*, *R. fuscus*, *Retiboletus* sp. HKAS 52680) are represented by China, five (*R. ornatipes*, *R. retipes* 1 & 2, *R. griseus*, *Retiboletus* sp. AF456813) are represented by USA, one (*R. nigerrimus*) from Japan and one (*R. flavoniger*) from Costa Rica.

Like the analysis of Zeng *et al.*, (2016) our analysis also shows polyphyletic existence of *R. retipes* (American species showing as “*R. retipes* 1” and “*R. retipes* 2” respectively in Figure 1). Present Indian collection (DC 16-58) is recovered here as sister to the Chinese *R. kauffmanii* with very high support (100% BS). The 100% bootstrap value supports the monophyly of *R. kauffmanii* + DC 16-58. In spite of our repeated efforts we failed to get good ITS sequence which further could give addition support to this interesting species.

Taxonomy

Retiboletus kauffmanii (Lohwag) N.K. Zeng & Zhu L. Yang; Mycologia 108, 2016 Figures 2 & 3.

Pileus 45–62 mm diam, subhemispherical to convex, sometimes helmet-shaped; surface dry, with blunt warts (verrucae) or blister-like protuberances, greyish brown (5D3) or brown (5E4), olive yellow (3D7) with NH₄OH and KOH and unchanging with FeSO₄ on pileus surface; margin with sterile flap of tissue. Pore surface light yellow to yellow (2A5–2A6), slightly darker when bruised; pores 0.8 per mm, rounded to somewhat angular, simple to compound. Tubes 5–7 mm long, subdecurrent to decurrent, concolorous to pore surface. Stipe 95–120 × 20–30 mm, central, cylindrical, gradually broader towards base; surface distinctly covered with reticulation (covering 1/3 to 1/2 of the stipe length from apex) which gradually becoming indistinct towards base; light yellow (2A5), slightly paler towards base, brownish orange (5C6) with NH₄OH, reddish brown (8F8) with KOH and unchanging with FeSO₄. Context pastel yellow (1A4) at pileus, greenish yellow (1A7 and 1A8) with NH₄OH and KOH; pastel yellow (2A4) or darker at stipe but yellowish orange (4A6) at base. Odor indistinct. Spore print olive-brown (4D4).

Research Article

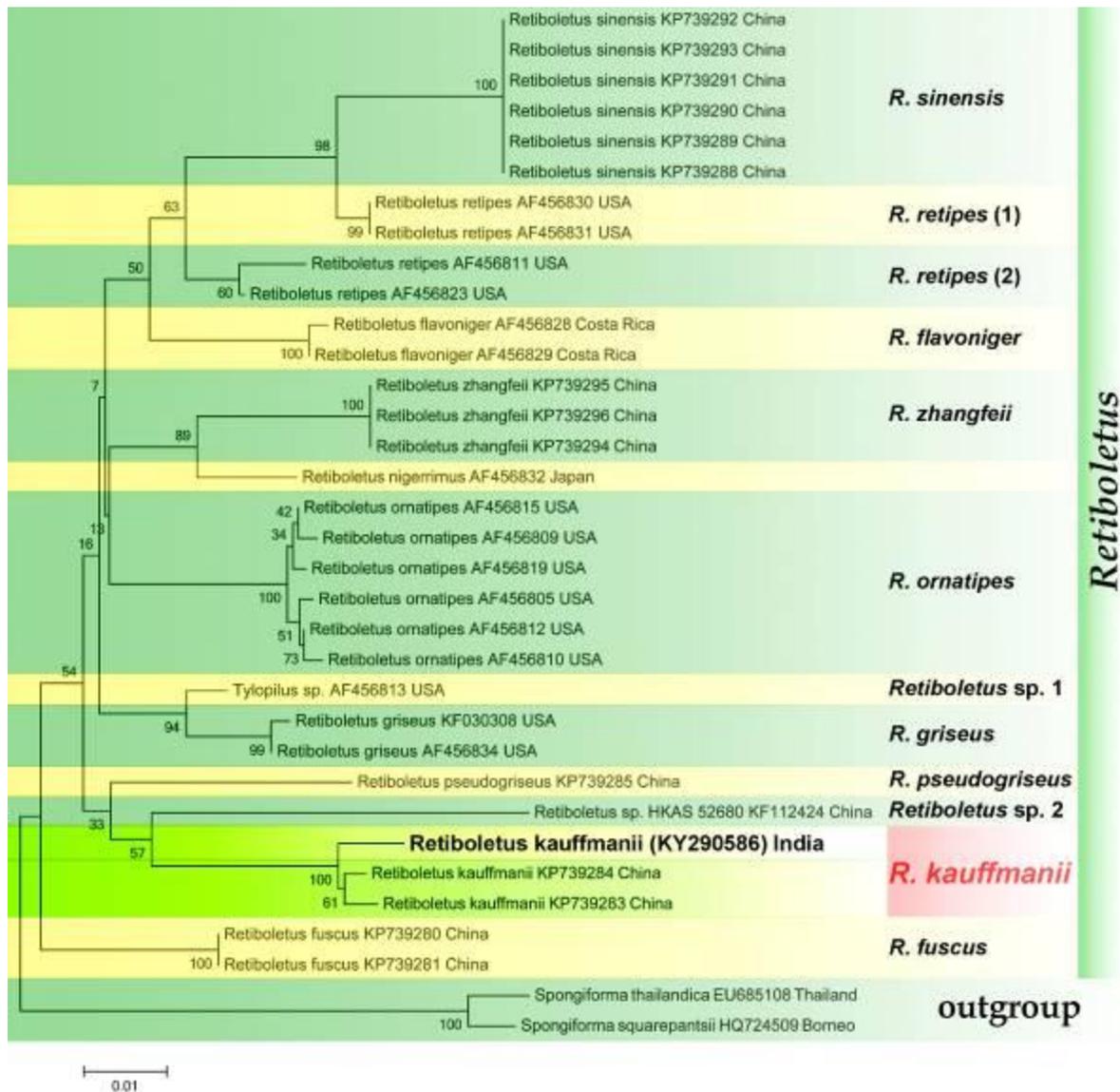


Figure 1: Phylogeny of DC 16-58 (Indian Collection of *Retiboletus Kauffmanii*, is in Bold and Black Font) Inferred from Maximum Likelihood Analysis of nrLSU Sequences Using MEGA 6.0

Basidiospores $9.9\text{--}11.8\text{--}13 \times 4.3\text{--}4.6\text{--}5.1 \mu\text{m}$ ($n = 20$, $Q = 2.25\text{--}2.53\text{--}2.79$), oblong, inequilateral, smooth under light microscope, olivaceous brown. Basidia $28\text{--}44 \times 11\text{--}15 \mu\text{m}$, 4-spored, clavate to subclavate. Pleurocystidia $45\text{--}60 \times 8\text{--}12 \mu\text{m}$, common, subfusiform to ventricose; content dense, granular, yellowish brown in KOH. Tube edge fertile, composed of basidia and cheilocystidia. Cheilocystidia shorter than pleurocystidia, $34\text{--}50 \times 7\text{--}11 \mu\text{m}$, cylindrical to subventricose. Hymenophoral trama divergent. Pileipellis $180\text{--}250 \mu\text{m}$ thick, a trichodermium, composed of more or less vertically arranged, sometimes interwoven, olive brown pigmented, highly encrusted hyphae; terminal cells $30\text{--}100 \times 10\text{--}16 \mu\text{m}$, cylindrical to subfusoid, with rounded apex, no color change in Melzer's reagent; incrustations in the form of rings or belts placed in parallel to subparallel tiers. Stipitipellis $40\text{--}60 \mu\text{m}$ thick, fertile, composed of caulobasidia, caulocystidia and marginal cells. Caulobasidia $30\text{--}46 \times 9\text{--}12 \mu\text{m}$, clavate, 2- to 4-spored. Caulocystidia $20\text{--}63 \times 8\text{--}13 \mu\text{m}$, subfusiform to fusiform, ventricose or appendiculate, mostly brown pigmented, content granular. Clamp connections absent in all tissues.

Research Article

Habitat/Distribution:—Gregarious, under *Lithocarpus* sp., in temperate broadleaf forest (dominated by *Lithocarpus*, *Castanopsis* and *Quercus*) of Sikkim.

Specimen Examined:—India: Sikkim, East Distr., Maenam Top, alt. 2315 m, N27°19'18.7'' E88°22'07.9'', 21 Aug. 2016, K. Das & D. Chakraborty, DC 16-058.



Figure 2: *Retiboletus kauffmanii* (DC 16-58); A & B: Fresh and Dissected Basidiomata; C: Pore Surface; D: Transverse Section through Pileipellis; E & F: Heavily Encrusted Hyphae of Pileipellis; G: Caulocystidia; H: Pleurocystidia; I & J: Transverse Section through Stipitipellis; K & L: Basidiospores in Phloxin and KOH Respectively; Scale bars: D = 100 µm; E–L = 10 µm

Research Article

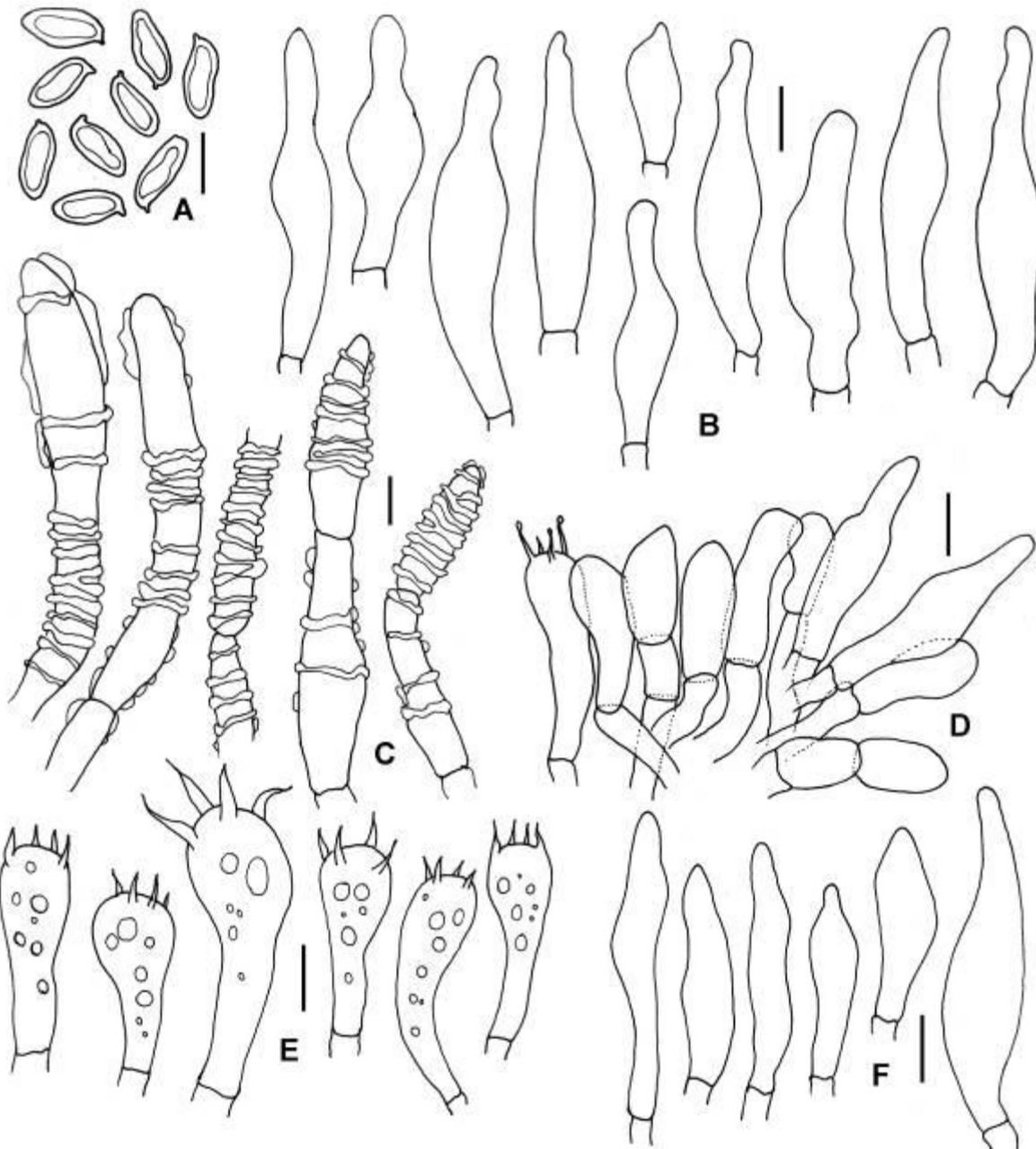


Figure 3: *Retiboletus kauffmanii* (Drawings from DC 16-58); A: Basidiospores; B: Caulocystidia; C: Terminal and Subterminal Cells of Heavily Encrusted Hyphae in Pileipellis; D: Transverse Section through Stipitipellis; E: Tube Basidia; F: Cheilocystidia; Scale Bars: A–F = 10 μ m

Notes: Macro- and micro-morphology of the present Indian collection mostly agree with that of Chinese specimens of *R. kauffmanii* except the prominent verrucoid pilear surface in our specimen. But, these verrucae might be either anomalous growth or well be of teratological origin. Present description is based on two basidiomata from a single collection site. So, additional collections are required to confirm the stability of this macro-morphological feature. Micro-morphologically, unlike the Chinese material Indian collection possesses encrusted hyphae (in pileipellis) which could be considered as a diagnostic feature for an undescribed species if the phylogeny is allowed us to do so. Our nrLSU-based analysis is not adequate to discriminate it up to a species rank, rather it indicate the conspecificity of Indian and Chinese

Research Article

collections. Until more field numbers are examined, those features can be considered here as morphological variations within *R. kauffmanii* only.

Retiboletus sinensis N.K. Zeng & Zhu L. Yang (represented by KP739288 to KP739293 in Figure 1) is morphologically close to *R. kauffmanii* but, differs by absence of wart or blister-like numerous projections on pileus surface, presence of yellow pores which turns brownish when injured, reticulations on stipe surface that extended nearly up to base and hyphal elements of pileipellis without any ring like incrustations and smaller basidiospores (8–10 × 3.5–4 µm) as mentioned in Zeng *et al.*, (2016).

ACKNOWLEDGEMENTS

We the authors are thankful to the Director, Botanical Survey of India (BSI), Kolkata and Scientist-in-charge, BSI, Gangtok for providing facilities to undertake this study. The forest department of the Government of Sikkim is acknowledged allowing DC & KD to undertake macrofungal surveys to South and East districts of Sikkim. SA, JK and TKB would like to acknowledge DST-PURSE and DST-FIST for providing central instrumentation facilities.

REFERENCES

- Binder M and Bresinsky A (2002).** *Retiboletus*, a new genus for a species-complex in the Boletaceae producing retipolides. *Feddes Repertorium* **113** 30–40.
- Crous PW, Gams W, Stalpers JA, Robert V and Stegehuis G (2004).** MycoBank: An online initiative to launch mycology into the 21st century. *Studies in Mycology* **50** 19–22.
- Desjardin DE, Binder M, Roekring S and Flegel T (2009).** *Spongiforma*, a new genus of gasteroid boletes from Thailand. *Fungal Diversity* **37** 1–8.
- Katoh K, Misawa K, Kuma K and Miyata T (2002).** MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Research* **30** 3059–3066.
- Kornerup A and Wanscher JH (1978).** *Methuen Handbook of Color*, 3rd edition, (Eyre Methuen Ltd., London, UK).
- Li GJ, Hyde KD, Zhao RL, Sinang H, Abdel-Aziz FA, Abdel-Wahab MA, Alvarado P, Alves-Silva G, Ammirati J, Ariyawansa HA, Baghela A, Bahkali AH, Beug M, Bhat DJ, Bojantchev D, Boonpratuang T, Bulgakov T, Camporesi E, Boro MC, Ceska O, Chakraborty D, Chen JJ, Chethana KWT, Chomnunti P, Consiglio G, Cui BK, Dai D, Dai YC, Daranagama DA, Das K, Dayarathne MC, Crop ED, De Oliveira RJV, Fragoso de Souza CA, Ivanildo de Souza J, Dentinger BTM, Dissanayake AJ, Doilom M, Drechsler-Santos ER, Ghobad-Nejhad M, Gilmore SP, Góes-Neto A, Gorczak M, Haitjema CH, Hapuarachchi KK, Hashimoto A, He MQ, Henske JK, Hirayama K, Iribarren MJ, Jayasiri SC, James TY, Jayawardena RS, Jeon SJ, Jerônimo GH, Jesus AL, Jones EBG, Kang JC, Karunarathna SC, Kirk PM, Konta S, Kuhnert E, Langer E, Lee HS, Lee HB, Li WJ, Li XH, Liimatainen K, Lima DX, Lin CG, Liu JK, Liu X, Liu ZY, Luangsa-ard JJ, Lücking R, Lumbsch HT, Lumyong S, Maharachchikumbura SSN, Leñaño EM, Marano AV, Matsumura M, McKenzie EHC, Mongkolsamrit S, Mortimer P, Nguyen TTT, Niskanen T, Norphanphoun C, O'Malley MA, Parnmen S, Pawłowska J, Perera RH, Peršoh D, Philips A, Phookamsak R, Phukhamsakda C, Pires-Zottarelli CLA, Raspé O, Reck MA, Cristina de Oliveira Rocha S, Monteiro de Azevedo Santiago ALC, Senanayake IC, Setti L, Shang QJ, Singh SK, Sir EB, Solomon KV, Song S, Srikitikulchai P, Stadler M, Suetrong S, Takahashi H, Takahashi T, Tanaka K, Tang LP, Thambugala KM, Thanakitpipattana D, Theodorou MK, Thongbai B, Thummarukcharoen T, Tian Q, Tibpromma S, Verbeken A, Vizzini A, Vlasák J, Voigt K, Wanasinghe DN, Wang Y, Weerakoon G, Wen HA, Wen TC, Wijayawardene NN, Wongkanoun S, Wrzosek M, Xiao YP, Xu J, Yan JY, Yang J, Yang SD, Yu H, Zhang JF, Zhao J and Zhou LW (2016).** Fungal diversity notes 253–366: taxonomic and phylogenetic contributions to fungal taxa. *Fungal Diversity* **78** 1-237 doi 10.1007/s13225-016-0366-9.
- Nuhn ME, Binder M, Taylor AFS, Halling RE and Hibbett DS (2013).** Phylogenetic overview of the Boletineae. *Fungal Biology* **117** 479–511.

Research Article

Tamura K, Stecher G, Peterson D, Filipski A and Kumar S (2013). MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution* **30** 2725–2729.

Thiers B [continuously updated]. *Index Herbariorum: A Global Directory of Public Herbaria and Associated Staff.* New York Botanical Garden's Virtual Herbarium. Available: <http://sweetgum.nybg.org/science/ih/>.

Vilgalys R and Hester M (1990). Rapid genetic identification and mapping enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* **172** 4238–4246.

Wu G, Feng B, Xu J, Zhu XT, Li YC, Zeng NK, Hosen MI and Yang ZL (2014). Molecular phylogenetic analyses redefine seven major clades and reveal 22 new generic clades in the fungal family Boletaceae. *Fungal Diversity* **69** 93–115.

Wu G, Li Y-C, Zhu X-T, Zhao K, Han L-H, Cui Y-Y, Li F, Xu J-P and Yang ZL (2016). One hundred noteworthy boletes from China. *Fungal Diversity* **81**(1) DOI 10.1007/s13225-016-0375-8.

Zeng NK, Liang ZQ, Wu G, Li YC, Yang ZL and Liang ZQ (2016). The genus *Retiboletus* in China. *Mycologia* **108** 363–380.