Case Report

NEW INSIGHTS INTO THE MECHANISM OF RAPHE NUCLEUS CONTROL OF COGNITION AND BEHAVIOUR: INTERACTION WITH DENDRITIC AND T CELLS

^{*}Manoj G Tyagi and Anand R

*Department of Pharmacology, Christian Medical College Vellore 632002 Tamilnadu, India *Author for Correspondence

ABSTRACT

The raphe nucleus in the brain i.e dorsal and median raphe nucleus is the main source of serotonin (5-HT), an important brain chemical in regulating cognition and behavior. The raphe nucleus is well connected neuronally to medial prefrontal cortex, limbic system and also hippocampus. Signal integration in pyramidal neurons is exerted at various cellular levels, with a key role played by the large apical dendrites. These are highly enriched in serotonergic receptors. There is now evidence showing a direct pathway from the retina to the raphe nucleus suggesting that optic stimulation may directly influence dorsal raphe nucleus neurons (DRN). 5-HT receptors located on optic afferent terminals can exert presynaptic inhibition of retinocollicular input and direct electrical stimulation of raphe nucleus. Serotonin in the DRN exhibits a diurnal rhythm that is influenced by sleep waking cycle and light dark cycle. On the other hand, Dendritic cells (DC) are capable of activating T cells. DCs take up and sequester 5-HT in lysosomal vesicles for subsequent release. Because DCs are specialized to stimulate naive T cells and 5-HT is postulated to be taken up by T cells, 5-HT released from DCs may modulate T-cell function. It appears that sequestration of 5-HT from neurons to DC and T cells and vice versa following optical stimulation of raphe nucleus leads to a regulatory effect on cognition and behaviour.

Key Words: Cognition, Raphe Nucleus, Pre-Frontal Cortex, Serotonin, Dendritic Cells, T Cells

INTRODUCTION

The raphe nuclei have a vast impact upon the central nervous system (Adell et al., 2002). Many of the neurons in the nuclei are serotonergic; i.e., contain 5-HT, a type of monoamine neurotransmitter. The raphe nuclei are distributed near the midline of the brainstem along its entire rostro-caudal extension. The serotonergic neurons are their main neuronal components, although a proportion of them lie in subdivisions of the lateral reticular formation. They develop from mesopontine and medullary primordia, and the resulting grouping into rostral and caudal clusters is maintained into adulthood, and is reflected in the connectivity. The dorsal raphe nucleus (DRN) of the mesencephalon is a complex multi-functional and multi-transmitter nucleus involved in a wide range of behavioral and physiological processes. Numerous studies demonstrate that the DRN receives a wide range of inputs including afferents from the locus coeruleus, the lateral habenula, several midbrain areas including the substantia nigra, and the periaqueductal gray, as well as fibers from the hypothalamus and the medial prefrontal cortex. The retina sends axons to the DRN which is a retino-raphe projection. Previous studies showed that the DRN received a direct retinal input, which consisted of a small number of retinal ganglion cells (RGCs), some of which exhibited alpha-like morphology (Foote et al., 1978 and Shen and Samba, 1994). Fite and colleagues continued this line of work and reported a substantial number of DRN-projecting RGCs, with both small and large soma sizes (Fite et al., 1999) and suggested that these cells arose from the non-image forming component of the retina (Fite et al., 2003). Intrinsically photosensitive retinal ganglion cells (ipRGCs) are considered the primary retinal component mediating non-image forming functions and these cells project to various visual and non-visual nuclei including lateral geniculate nucleus (LGN) (Provencio et al., 2002 and Hattar et al., 2002) suprachiasmatic nucleus (SCN), intergeniculate leaflet

Case Report

(IGL) of the LGN complex, and olivary pretectal nucleus (Morin *et al.*, 2003; Sollars *et al.*, 2003; Dacey *et al.*, 2005 and Baver *et al.*, 2008). In this review we evaluate the influence of retina on raphe nuclear levels of 5-HT and its sequestration into DC and T cells.

General Anatomical and Functional Characteristics of 5ht System

The 5-HT-producing neurons are mainly located in the brainstem raphe nuclei that have been shown to give rise to two major groups of neurons: (1) the superior group at the interface between the midbrain and the pons; and (2) the inferior group located more caudally in the pons (Azmitia *et al.*, 1995). They form the largest and complex neuro-chemical efferent system in the brain. The superior group of 5-HT neurons comprising the dorsal and median raphe nuclei is the source of projections to various sites in the forebrain as shown in Figure1.



Figure 1: Neuronal pathways of raphe nucleus in the brain

Rich 5-HT innervations of telencephalic limbic regions such as the prefrontal and cingulate cortices, the amygdala, hippocampus, and ventral striatum, and diencephalic structures, especially the hypothalamus and thalamus, are found (Azmitia et al., 1995 and Bentivoglio et al., 1993). The dorsal and median raphe nuclei differentially innervate the forebrain regions. For instance, the dorsal raphe nucleus provides projections primarily to the amygdala and ventral striatum, whereas the median raphe nucleus preferentially innervates the prefrontal and cingulate cortices and the hippocampus. The least levels of 5-HT fibers are seen in the motor regions of the frontal lobe (Murphy et al., 1998). The inferior group of 5HT-containing neurons sends abundant descending spinal projections. Therefore, determining the morphological and physiological properties of DRN-projecting RGCs will provide much needed information about the type of retinal information processing performed by the DRN. The prefrontal cortex is involved in an array of higher brain functions that are altered in psychiatric disorders. Serotonergic neurons of the midbrain raphe nuclei innervate the prefrontal cortex and are the cellular target for drugs used to treat mood disorders such as the selective serotonin (5-HT) reuptake inhibitors (Abi-Saab et al., 1999). Anatomical evidence supports the existence of projections from the medial prefrontal cortex (mPFC) to the dorsal raphe nucleus (DR). Pyramidal neurons of the mPFC co-express postsynaptic 5-HT (1A) (inhibitory) and 5-HT (2A) (excitatory) receptors. Consistent with the above observations, the selective activation of both receptors in mPFC reduced and increased, respectively, the firing activity of DR 5-HT neurons and the 5-HT release in mPFC. Overall, these data indicate that the activity of the 5-HT

Case Report

system is strongly controlled by the mPFC. Moreover, the activation of postsynaptic 5-HT1A receptors in the mPFC reduced the local release of 5-HT (Casanovas *et al.*, 1999) and the firing rate of dorsal raphe 5-HT neurons (Birkett and Fite, 2005). These data suggest that pyramidal neurons containing 5-HT 1A receptors may play a role in the distal control of serotonergic activity. The 5-HT system has been widely demonstrated to be involved in the pathogenesis of diverse mental illnesses such as obsessive compulsive disorder (OCD). Several lines of evidence suggest that the dysfunction of 5-HT neurotransmission, and especially an altered sensitivity of the 5-HT receptor subtype, may constitute a crucial factor in the patho-physiology of OCD.

Immune System and Central Nervous System

Immune cell function in the CNS has now been shown to extend beyond pathological conditions. Indeed, recent data have suggested key roles for immune cells in healthy brain functions, including psychological stress responses, spatial learning and memory, and adult neurogenesis (Kipnis et al., 2004; Brynskikh et al., 2008; Ziv et al., 2006 and Goehler et al., 1999). In reality there is abundant communication between the immune system and the CNS. For example, intraperitoneal injection of pro-inflammatory cytokines was shown to generate CNS-mediated sickness behaviour, which could be blocked by vagus nerve transaction (Akwa et al., 1998). The beneficial effect of T cells specific for CNS-restricted self antigens has been observed in models of optic nerve injury, spinal cord contusion and stroke, as well as in other models of acute and chronic neurodegenerative conditions. T cells have been proposed to mediate their neuroprotective effect via the production of neurotrophins, the modulation of glutamate release by astrocytes and microglia, the regulation of innate immunity at the site of injury and other, as yet unexplored, mechanisms. These data suggest that there is a link between the neuroprotective function of T cells and their recognition of self antigens. The possible contribution of astrocytes to immune responses within the brain has been described in several settings, including those involving the targeted overexpression of cytokines such as TNF, IFN- α , TGF- β , IL-6, and IL-12 by astrocytes, which leads to chronic inflammation and progressive neurodegeneration (Pagenstecher et al 2000, Wyss-Coray et al 1997, Krishnamoorthy et al., 2007). More recent studies analyzing mice in which the ability of astrocytes to participate in immune function is compromised through the specific loss of a cytokine receptor such as gp130 or reduced NF- κ B signaling, have shown that this alters the course of immune responses in the CNS (Drogemuller et al., 2008). Thus, in a mouse model of spinal cord injury, astrocyte-specific inhibition of NF-kB (which is necessary for the activation of many cytokine genes) resulted in a reduction in the number of reactive astrocytes in the CNS, in lower levels of chemokines, and in reduced infiltration of T cells and macrophages (Sofroniew, 2005). Consequently, this led to improved spinal cord healing. Future challenges include determining how individual cytokines, adhesion molecules, and chemokines produced by astrocytes influence the development of inflammation and the behavior of infiltrating immune cell populations.

Sequestration of 5-Ht from the Raphe Nucleus into Immune Cells and Vice Versa and Distribution into Discrete Brain Sites: A Novel Hypothesis

Several types of immune cells including B and T cells, granulocytes, macrophages, mast cells and dendritic cells are located within the meningeal structures of the brain. Although functional roles of these DRN-projecting ganglion cells remains unclear, there is evidence that DRN neurons respond to changes in the light and dark cycle (Nautiyal *et al.*, 2011 and Wolf *et al.*, 2009) and they are sensitive to phasic flashing light stimulation. There is a direct anatomical connection between the DRN and SCN (Filippova *et al.*, 2004 and Heym *et al.*, 1982). Furthermore, in addition to the conventional retino-hypothalamic tract (RHT) that provides luminance information necessary for entrainment, brief millisecond photo stimulation has been shown to be capable of inducing circadian phase shifts (Van Den Pol *et al.*, 1998 and Morin, 1999). All the members of the 5HT1 receptor subtype belong to the family of G protein-coupled receptors. They generally reduce adenylate cyclase activity, leading to decreased cyclic adenosine monophosphate (cAMP) production. The 5TH1A receptor represents a somato-dendritic autoreceptor on

Case Report

the cell body of 5HT neurons in the brainstem raphe nuclei. Another subtype, the 5HT1B receptor and its human homolog, 5HT1D has been found to function as an autoreceptor on axon terminals (Arvanitogiannis and Amir, 1999). When activated, these two receptors attenuate the intrinsic firing of the raphe cells, thereby inhibiting 5HT release. The 5HT1A receptors have also been characterized at postsynaptic sites (Backstrom *et al.*, 1995 and Marek and Aghajanian, 1999). A significant amount of 5HT1B receptors are present on postsynaptic structures, although their function is still unknown (Sander-Bush and Mayer, 2001). Therefore, there could be a second pathway that conveys fast luminance changing signals to SCN. Dendritic cells (DC) are capable of activating T cells. DCs take up and can sequester 5-HT in lysosomal vesicles for subsequent release. Because DCs are specialized to stimulate naive T cells and 5-HT is postulated to be taken up by T cells, 5-HT released from DCs may modulate T-cell function (Eberl *et al.*, 2004). It appears that sequestration of 5-HT from neurons to DC and T cells following optical stimulation of raphe nucleus leads to a regulatory effect on memory and learning (Hornung, 2003 and Tyagi, 2012). The effect of T cells on the CNS might also be mediated via soluble cytokines that are released into the circulation. This raises the issue of the variability of blood–brain barrier permeability and how this influences the possibility of a peripheral T cell effect (Bird, 2005).

CONCLUSION

This study clearly suggests a novel mechanism for 5-HT sequestration and distribution in the brain after photo-stimulation. There is enough evidence showing a direct pathway from the retina to the raphe nucleus suggesting that optical stimulation may directly influence dorsal raphe nucleus neurons (DRN). 5-HT receptors located on optic afferent terminals can exert pre-synaptic inhibition of retinocollicular input and direct electrical stimulation of raphe nucleus. It is suggested that DCs take up and sequester 5-HT in lysosomal vesicles for subsequent release. Because DCs are specialized to stimulate naive T cells and 5-HT is postulated to be taken up by T cells, 5-HT released from DCs may modulate T-cell function. It appears that sequestration of 5-HT from neurons to DC and T cells and vice versa following optical stimulation of raphe nucleus leads to a regulatory effect on cognition and behavior.

ACKNOWLEDGEMENT

The author is thankful to Dr.Adrian C Hayday, King's College, London for useful scientific discussions.

REFERENCES

Adell A, Celada P, Abellán MT and Artigas F (2002). Origin and functional role of the extracellular serotonin in the midbrain raphe nuclei. *Brain Research Reviews* **39** 154-180.

Foote WE, Taber-Pierce E and Edwards (1978). Evidence for a retinal projection to the midbrain raphe of the cat. *Brain Research* 156 135-140.

Shen H and Semba K (1994). A direct retinal projection to the dorsal raphe nucleus in the rat. *Brain Research* 635 159-168.

Fite KV, Janusonis S, Foote WE and Bengston L (1999). Retinal afferents to the dorsal raphe nucleus in rats and Mongolian gerbils. *Journal of Comparative Neurology* **414** 469-484.

Fite KV, Birkett MA, Smith A, Janusonis S and McLaughlin S (2003). Retinal ganglion cells projecting to the dorsal raphe and lateral geniculate complex in Mongolian gerbils. *Brain Research* 973 146-150.

Provencio I, Rollag MD and Castrucci AM (2002). Photoreceptive net in the mammalian retina. This mesh of cells may explain how some blind mice can still tell day from night. *Nature* **415** 493.

Hattar S, Liao HW, Takao M, Berson DM and Yau KW (2002). Melanopsin-containing retinal ganglion cells: architecture, projections, and intrinsic photosensitivity. *Science* 295 1065-1070.

Case Report

Morin LP, Blanchard JH and Provencio I (2003). Retinal ganglion cell projections to the hamster suprachiasmatic nucleus, intergeniculate leaflet, and visual midbrain: bifurcation and melanopsin immunoreactivity. *Journal of Comparative Neurology* **465** 401-416.

Sollars PJ, Smeraski CA, Kaufman JD, Ogilvie MD and Provencio I (2003). Melanopsin and nonmelanopsin expressing retinal ganglion cells innervate the hypothalamic suprachiasmatic nucleus. *Visual Neuroscience* 20 601-610.

Dacey DM, Liao HW, Peterson BB, Robinson FR and Smith VC (2005). Melanopsin-expressing ganglion cells in primate retina signal colour and irradiance and project to the LGN. *Nature* **433** 749-754.

Baver SB, Pickard GE, Sollars PJ and Pickard GE (2008). Two types of melanopsin retinal ganglion cell differentially innervate the hypothalamic suprachiasmatic nucleus and the olivary pretectal nucleus. *European Journal of Neuroscience* 7 1763-1770.

Brown TM, Gias C, Hatori M, Keding SR and Semo M (2010). Melanopsin contributions to irradiance coding in the thalamo-cortical visual system. *PLOS Biology* **8** e1000558.

Azmitia EC, Rubinstein VJ, Strafaci JA, Rios JC and Whitaker - Azmitia PM (1995). 5-HT1A agonist and dexamethasone reversal of para-chloroamphetamine induced loss of MAP-2 and synaptophysin immunoreactivity in adult rat brain. *Brain Research* 677(2) 181-192.

Bentivoglio M, Kultas-Ilinsly K and Ilinsly I (1993). Limbic thalamus: structure, intrinsic organization, and connections. In: *Neurobiology of cingulate cortex and limbic thalamus: a comprehensive handbook* Edited by Vogt BA, Gabriel M, Boston: Birkhäuser 71-122.

Murphy DL, Andrews AM, Wichems CH, Li Q, Tohda M and Greenberg B (1998). Brain serotonin neurotransmission: an overview and update with an emphasis on serotonin subsystem heterogeneity, multiple receptors, interactions with other neurotransmitter systems, and consequent implications for understanding the actions of serotonergic drugs. *Journal of Clinical Psychiatry* **59**(Suppl 15) 4-12.

Abi-Saab WM, Bubser M, Roth RH and Deutch AY (1999). 5-HT2 receptor regulation of extracellular GABA levels in the prefrontal cortex. *Neuropsychopharmacology* **20**(1) 92-96.

Casanovas JM, Hervás I and Artigas F (1999). Postsynaptic 5-HT1A receptors control 5-HT release in the rat medial prefrontal cortex. *Neuroreport* **10**(7) 1441-1445.

Birkett M and Fite KV (2005). Diurnal variation in serotonin immunoreactivity in the dorsal raphe nucleus. *Brain Research* 1034 180-184.

Kipnis J, Cohen H, Cardon M, Ziv Y and Schwartz M (2004). T cell deficiency leads to cognitive dysfunction: implications for therapeutic vaccination for schizophrenia and other psychiatric conditions. *Proceedings of the National Academy of Sciences USA* **101** 8180-8185.

Brynskikh A, Warren T, Zhu J and Kipnis J (2008). Adaptive immunity affects learning behavior in mice. *Brain Behavior and Immunity* 22 861-869.

Ziv Y, Ron N, Butovsky O, Landa G, Sudai E, Greenberg E, Cohen H, Kipnis J and Schwartz M (2006). Immune cells contribute to the maintenance of neurogenesis and spatial learning abilities in adulthood. *Nature Neuroscience* 9 268-275.

Goehler, Ron LE, Gaykema PA, Kien T Nguyen, Jacqueline E Lee, Fred JH Tilders, Steven F Maier and Linda R Watkins (1999). Interleukin1 β in immune cells of the abdominal vagus nerve: a link between the immune and nervous systems. *Journal of Neuroscience* **19** 2799-2806.

Akwa Y, Hassett DE, Eloranta ML, Sandberg K, Masliah E, Powell H, Whitton JL, Bloom FE and Campbell (1998). Transgenic expression of IFN-alpha in the central nervous system of mice protects against lethal neurotropic viral infection but induces inflammation and neurodegeneration. *Journal of Immunology* 161(9) 5016-5026.

Pagenstecher A, Lassmann S, Carson MJ, Kincaid CL, Stalder AK and Campbell IL (2000). Astrocyte-targeted expression of IL-12 induces active cellular immune responses in the central nervous system and modulates experimental allergic encephalomyelitis. *Journal of Immunology* **164**(9) 4481-4492.

Case Report

25. Wyss-Coray T, Borrow P, Brooker MJ and Mucke L (1997). Astroglial overproduction of TGFbeta 1 enhances inflammatory central nervous system disease in transgenic mice. *Journal of Neuroimmunology* 77(1) 45-50.

Krishnamoorthy G, Holz A and Wekerle H (2007). Experimental models of spontaneous autoimmune disease in the central nervous system. *Journal of Molecular Medicine* **85**(11) 1161-1173.

Drögemüller K, Helmuth U, Brunn A, Sakowicz-Burkiewicz M, Gutmann DH, Mueller W, Deckert M and Schlüter D (2008). Astrocyte gp130 expression is critical for the control of Toxoplasma encephalitis. *Journal of Immunology* **181**(4) 2683-2693.

Sofroniew MV (2005). Reactive astrocytes in neural repair and protection. *Neuroscientist* **11**(5) 400-407. **Nautiyal KM, Liu C, Dong X and Silver R (2011).** Blood-borne donor mast cell precursors migrate to

mast cell-rich brain regions in the adult mouse. *Journal of Neuroimmunology* 240-241 and 142-146. Wolf SA Steiner B Wengner A Linn M Kammertoens T Kempermann G (2000) Adaptive

Wolf SA, Steiner B, Wengner A, Lipp M, Kammertoens T, Kempermann G (2009). Adaptive peripheral immune response increases proliferation of neural precursor cells in the adult hippocampus. *FASEB Journal* 23 3121-3128.

Filippova IV, Williams WC and Frolov VA (2004). Very slow potential oscillations in locus coeruleus and dorsal raphe nucleus under different illumination in freely moving rats. *Neuroscience Letters* **363** 89-93.

Heym J, Trulson ME and Jacobs BL (1982). Raphe unit activity in freely moving cats: effects of phasic auditory and visual stimuli. *Brain Research* 232 29-39.

Van den Pol AN, Cao V and Heller HG (1998). Circadian system of mice integrates brief light stimuli. *American Journal of Physiology - Regulatory, Integrative and Comparative Physiology* **275** R654-R657.

Morin LP (1999). Serotonin and the regulation of mammalian circadian rhythmicity. *Annals of Medicine* **31** 12-33.

Arvanitogiannis A and Amir S (1999). Resetting the rat circadian clock by ultrashort light flashes. *Neuroscience Letters* 261 159-162.

Backstrom JR, Westphal RS, Canton H and Sanders-Bush E (1995). Identification of rat serotonin 5-HT2c receptors as glycoproteins containing N-linked oligosaccharides. *Brain Research Molecular Brain Research* **33**(2) 311-318.

Marek GJ and Aghajanian GK (1999). 5-HT2A receptor or alpha1-adrenoceptor activation induces excitatory postsynaptic currents in layer V pyramidal cells of the medial prefrontal cortex. *European Journal of Pharmacology* 367 197-206.

Sanders-Bush E and Mayer SE (2001). 5-hydroxytryptamine (serotonin): receptor agonists and antagonists. In: Hardman JG, Limbird LE, Goodman Gilman A, editors. *Goodman and Gilman's the Pharmacological Basis of Therapeutics*. 10. New York: McGraw-Hill 269-290.

Vidal L and Morin LP (2007). Absence of normal photic integration in the circadian visual system: response to millisecond light flashes. *Journal of Neuroscience* 27 3375-3382.

Eberl M, Jomaa H and Hayday AC (2004). Integrated immune responses to infection-cross-talk between human gamma delta Tcells and dendritic cells. *Immunology* **112**(3) 364-368.

Hornung JP (2003). The human raphe nuclei and the serotonergic system. *Journal of Chemical Neuroanatomy* 26(4) 331-343.

Tyagi MG (2012). New insights into the mechanism of raphe nucleus control of cognition. *Neurobiology of Cognition Symposium* NIMHANS Bangalore 61.

Bird L (2005). Immune responses: Shuttling serotonin not just in our heads. *Nature Reviews Immunology* 5 904.