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## **IMPACT OF SUPPLEMENTARY UV-B RADIATION ON THE MORPHOLOGY, GROWTH AND YIELD OF VIGNA MUNGO (L.) HEPPER VAR. ADT-3**

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### **ABSTRACT**

The effect of ultraviolet-B (UV-B) radiation on the morphology and growth of black gram *Vigna mungo* (L.) Hepper var. ADT-3 was evaluated. Screening of black gram after exposure to supplementary UV-B radiation (2 hours daily @ 12.2 kJ m<sup>-2</sup> d<sup>-1</sup>; ambient = 10 kJ m<sup>-2</sup> d<sup>-1</sup>) was carried out in terms of growth on 15, 30, 45 and 60 DAS. Supplementary UV-B irradiation reduced the number of leaves by 20 to 60 %. Total leaf area (83.23 %) and leaf area index (48.65 %). Specific leaf weight in UV-B irradiated increased by 18.08, 3.81 and 10.67 % on 15, 30 and 45 respectively, while it showed a reduction on 60 DAS. UV-B stress decreased the fresh weight of leaves by 7.93 to 80.74 % and dry weight of foliage by 33.33 to 76.08 % in all stages of UV-B exposed plants. Growth of all the varieties of black gram was progressively inhibited by the UV-B radiation. Suppression of root and shoot length ranged from 8.50 to 36.70 % respectively at all stages of growth resulting in reduced plant height and S/R (4.52 to 30.99 %). However, S/R ratio showed an increase of 38.91 % over control on 315 DAS. Plant dry weight after UV-B stress fell below control by 76.97 % 60 DAS. The relative growth rate (RGR) was reduced in all varieties with age as it reached 42.46 % reduction on 60 DAS. UV-B exposure decreased the entire yield components per plant basis, the decreases being 50 % in the pod number, 66.28 % in pod weight, 22.22 % in pod length, 61.29 % in seed number and 21.26 to 61.87 % in seed mass. Harvest index was the least under UV-B treatment which showed a reduction by 14.66 % compared with control. A similar pattern was obtained for data on shelling percentage also.

**Keywords:** Ultraviolet-B, Black Gram, var. ADT-3, Morphology, Growth, Yield

### **INTRODUCTION**

If a stress like ultraviolet-B (UV-B) radiation is imposed on the plant during the active growth phase, the impact would be reflected on the gross morphology mediated through the metabolic labyrinth, as the plant struggles to negotiate the adverse effects and perform normally (Rajendiran, 2001). Elevated ultraviolet-B severely damages leaves (Kokilavani and Rajendiran, 2013; Kokilavani and Rajendiran, 2014a; Kokilavani and Rajendiran, 2014b; Kokilavani and Rajendiran, 2014c), inhibits plant growth (Rajendiran and Ramanujam, 2003, Rajendiran and Ramanujam, 2004) and suppresses nodulation and nitrogen metabolism (Rajendiran and Ramanujam, 2006; Sudaroli and Rajendiran, 2013a; Sudaroli and Rajendiran, 2013b; Arulmozhi and Rajendiran, 2014; Vijayalakshmi and Rajendiran, 2014) in sensitive plants. Although literature on UV-B and plant interaction is voluminous, most of the works deal with the gross effects on growth and yield under controlled environmental conditions. Hardly 5% of the over 600 publications relate to field studies (Caldwell *et al.*, 1998). This has been blamed as the major defect leading to overstating the damaging influence of UV-B (Jordan, 1997) as plants under natural day light conditions with high PAR (photosynthetically active radiation) are affected very little (Adamse and Britz, 1992). The present work assumes further significance as the plants of black gram *Vigna mungo* (L.) Hepper var. ADT-3 has been grown under field conditions (*in situ*) allowing the interaction of PAR and other environmental factors.

### **MATERIALS AND METHODS**

Black gram (*Vigna mungo* (L.) Hepper var. ADT-3) the nitrogen fixing grain legume was chosen for the study. Viable seeds of *Vigna mungo* (L.) Hepper var. ADT-3 was procured from Saravana Farms,

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Villupuram, Tamil Nadu, India. The seeds were selected for uniform color, size and weight and used in the experiments. The crops were grown in pot culture in the naturally lit greenhouse (day temperature maximum  $38 \pm 2$  °C, night temperature minimum  $18 \pm 2$  °C, relative humidity  $60 \pm 5$  %, maximum irradiance (PAR)  $1400 \mu\text{mol m}^{-2} \text{s}^{-1}$ , photoperiod 12 to 14 h). Supplementary UV-B radiation was provided in UV garden by three UV-B lamps (*Philips TL20W/12 Sunlamps*, The Netherlands), which were suspended horizontally and wrapped with cellulose diacetate filters (0.076 mm) to filter UV-C radiation ( $< 280$  nm). UV-B exposure was given for 2 h daily from 10:00 to 11:00 and 15:00 to 16:00 starting from the 5th day after sowing. Plants received a biologically effective UV-B dose (UV-B<sub>BE</sub>) of  $12.2 \text{ kJ m}^{-2} \text{ d}^{-1}$  equivalents to simulated 20 % ozone depletion at Pondicherry ( $12^{\circ}2' \text{N}$ , India). The control plants, grown under natural solar radiation, received UV-B<sub>BE</sub>  $10 \text{ kJ m}^{-2} \text{ d}^{-1}$ . Ten plants from each treatment were carefully uprooted on 15, 30, 45 and 60 DAS and their axial growth (roots and shoot length and plant height) and fresh biomass were measured. They were then dried in an oven at 80° C for 48 h and weighed again for dry mass measurements. Alongside, morphological and developmental abnormalities if any, caused by UV-B radiation were also recorded. Assessment of growth of test plant on 15, 30, 45 and 60 DAS were recorded and calculated using standard methods. Ten plants were selected at random from each of the treatments. The leaf area (the leaflets from all the nodes) was determined at various stages using Area meter (Analytical Development Corporation, UK, model AM100). The total leaf area per plant was obtained by summing up the area of the leaves from all the nodes of the plant. Leaf area index (LAI) (Williams, 1946), specific leaf weight (SLW) (Pearce *et al.*, 1968), relative growth rate (RGR) (Williams, 1946) and shoot / root ratio (Racey *et al.*, 1983) were calculated using the following formulae.

$$\text{LAI} = \frac{\text{Leaf area of the plants (cm}^2\text{)}}{\text{Ground area occupied (cm}^2\text{)}}$$

$$\text{SLW} = \frac{\text{Leaf dry weight (g)}}{\text{Leaf area (m}^2\text{)}}$$

$$\text{RGR} = \frac{\text{Log}_e W_2 - \text{Log}_e W_1}{t_2 - t_1}$$

where,  $W_1$  and  $W_2$  are dry masses of whole plants at  $t_1$  and  $t_2$  (time in days) respectively.

$$\text{S/R ratio} = \frac{\text{Shoot weight (g)}}{\text{Root weight (g)}}$$

Mature fruits were harvested periodically from each plant and the length and weight of the pod, number of seeds per pod and number of seeds per plant and weight of seeds per plant were recorded. Harvest index (Mohan *et al.*, 1992) and shelling percentage (Francis *et al.*, 1978) were calculated using the following formulae.

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$$\text{Harvest index} = \frac{\text{Yield of the plant (g)}}{\text{Biomass of the plant (g)}} \times 100$$

$$\text{Shelling percentage} = \frac{\text{Seed wt. plant}^{-1}}{\text{Fruit wt. plant}^{-1}} \times 100$$

At least ten replicates were maintained for all treatments and control. The experiments were repeated to confirm the trends.

**RESULTS AND DISCUSSION**

In a series of experiments, the responses of black gram were assessed in terms of growth on 15, 30, 45 and 60 DAS. There were fewer leaves only (20 to 60 %) under UV-B stress on 45 and 60 DAS, but plants under normal ambience had more number of leaves (Table 1). Supplementary UV-B irradiation reduced the total leaf area throughout the growth period, the maximum being 83.23 % on 45 DAS (Table 1; Plate 1, Figure 1 to 4). The LAI was reduced by UV-B exposure to a larger extent (48.65 %) below control on 60 DAS (Table 1). The SLW in UV-B irradiated increased with age, showing a dip on 60 DAS (22.22 %). However, an increase of 18.08, 3.81 and 10.66 % were observed on 15, 30 and 45 DAS respectively. UV-B stress decreased the fresh weight of leaves by 7.93 to 80.74 %, with the maximum reduction being on 45 DAS. The dry weight of foliage decreased by 33.33 to 76.08 % in all stages of UV-B exposed plants (Table 1). Reductions in leaf area and mass were observed in the field-grown sweetgum plants (Sullivan *et al.*, 1994), black gram (Rajendiran and Ramanujam, 2000) and green gram (Rajendiran and Ramanujam, 2003) and ten varieties of cowpea (Kokilavani and Rajendiran, 2014d) exposed to elevated UV-B radiation. According to Britz and Adamse (1994) changes in the leaf area and dry mass indicated that cell elongations as well as cell contents were affected. On prolonged exposure to UV-B, the leaves of black gram exhibited various kinds of abnormalities. The leaves became generally pale which at times occurred in patches. The yellowing intensified and became discretely chlorotic (Plate 1, Figure 5). Browning developed in patches indicating necrosis of the underlying tissues during later stages. Necrotic lesions appeared in older leaves which have received UV-B over a long time (Plate 1, Figure 6). UV-B exposure reduced root length significantly on all stages of growth till 60 DAS. UV-B stressed plants showed maximum reduction of root growth 36.70 % on 15 DAS which recovered after 45 DAS (Table 2). Shoot length of UV-B stressed plants decreased by 8.50 % within 15 DAS and continued so till 60 DAS with 18.80 % reduction (Table 2). The S/R ratio was decreased by UV-B stress by 4.52 to 30.99 %. However, S/R ratio showed an increase of 38.91 % over control on 15 DAS (Table 2). Fresh weight of roots increased with age in all treatments. But the biomass accumulation in root was inhibited by UV-B treatment by 1 to 45.34 % on all stages of growth. The trends observed in root and shoot biomass pattern were reflected at the whole plant level too with inhibitions at UV-B, little improvement in later stages with maximum reduction of 76.97 % on 45 DAS (Table 2, Plate 1, Figure 7). Inhibition of growth indicated by reductions in root and shoot length and biomass content due to UV-B stress were apparent at all stages. Such inhibitions are characteristic of UV-B stressed legumes as in *Vigna unguiculata* (Kulandaivelu *et al.*, 1989), *Phaseolus vulgaris* (Mark and Tevini, 1997), *Vigna mungo* (Rajendiran and Ramanujam, 2000), *Vigna radiata* (Rajendiran and Ramanujam, 2003) and ten varieties of cowpea (Kokilavani and Rajendiran, 2014d). The stunting of UV-B stressed plants is attributed to destruction of endogenous IAA whose photo-oxidative products may be inhibitory (Kulandaivelu *et al.*, 1989; Tevini and Teramura, 1989) as indicated by a decrease in IAA content concomitant with a corresponding increase in IAA oxidase activity in rice leaves (Huang *et al.*, 1997). Total biomass represents a long-term

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integration of all biochemical, physiological and growth parameters. It is a good indicator of UV-B radiation effects on growth as low biomass appears to be an even response. Analyzed for the whole plant or plant parts, fresh or dry, shoot or root biomass accumulations or shoot / root ratios are often substantially reduced by UV-B (Nedunchezian and Kulandaivelu, 1997; Nogues *et al.*, 1998). Brandle *et al.*, (1977) found that the dry weight of pea was significantly reduced after nine days of UV-B irradiance. Similar results were obtained in *Phaseolus vulgaris* (Dumpert and Knacher, 1985), *Helianthus annuus* (Tevini *et al.*, 1991), *Oenothera stricta*, *Plantago lanceolata* and *Chamaesyce celastroides* (Ziska *et al.*, 1992), *Liquidamber styraciflua* (Sullivan *et al.*, 1994) and *Vigna radiata* (Rajendiran and Ramanujam, 2003) and in ten varieties of cowpea (Kokilavani and Rajendiran, 2014d).

Leaf biomass increased by UV-B irradiance while contribution of root and stem decreased (Jain *et al.*, 1999).

RGR was reduced with age as it reached 22.82 to 42.46 % reduction on 60 DAS (Table 2). Similar inhibitions of RGR by UV-B were observed by Jain *et al.*, (1999) in mungbean and Rajendiran, Ramanujam (2003) in green gram and in ten varieties of cowpea (Kokilavani and Rajendiran, 2014d).

Supplemental UV-B exposure consistently decreased the entire yield components per plant basis, the decreases being 50 % in the pod number, 66.28 % in pod weight, 22.22 % in pod length, 61.29 % in seed number and 21.26 to 61.87 % in seed mass (Table 3, Plate 1, Figure 9, 10). Analyzed on the basis of number of seeds per pod, only the UV-B treated plants had more fruits containing less number of seeds (Table 3).

Harvest index was the least under UV-B treatment which showed a reduction by 14.66 % compared with control. A similar pattern was obtained for data on shelling percentage also (Table 3). UV-B exposure delayed the flowering and reduced the yield in crop plants in general (Caldwell and Flint, 1994; Rajendiran and Ramanujam, 2004; Kokilavani and Rajendiran, 2014e).

Both the timing of flowering and the number of flowers produced in maize cultivars, *Petunia hybrida*, *Brassica rapa* and several mono- and dicotyledonous ephemerals have been altered by UV-B (Staxen and Bornman, 1994; Musil, 1995; Klaper *et al.*, 1996; Mark *et al.*, 1996, Rajendiran and Ramanujam, 2004). Since the success of pollination is linked to the availability of pollinators, any phenological dislocation would affect the reproductive success greatly (Caldwell, 1968; Ziska *et al.*, 1992; Staxen and Bornman, 1994; Musil, 1995; Klaper *et al.*, 1996; Mark *et al.*, 1996; Rajendiran and Ramanujam, 2004). In fact, the reproductive organs like pollen and ovules have effective protective features. The UV-B absorbing compounds are abundant in the floral parts (sepals and petals) and more specifically on the walls of pollens and ovaries (Day and Demchik, 1996).

Though very little work has been done on the phenological changes, even minor alterations could adversely affect the agricultural systems. Simulating a 40 % ozone reduction, Esser (1980) found the yield of potato reduced by 41 %, spinach by 66 %, cabbage by 49 % and bean by 75 %, for a 20 % ozone depletion the yield of green gram was decreased by 25 to 45 % (Rajendiran and Ramanujam, 2004) and 20 to 56 % reduction in cowpea (Kokilavani and Rajendiran, 2014e). In 1991, Giller observed the yield of cotton and soybean reduced by 23 and 25% respectively under field conditions also.

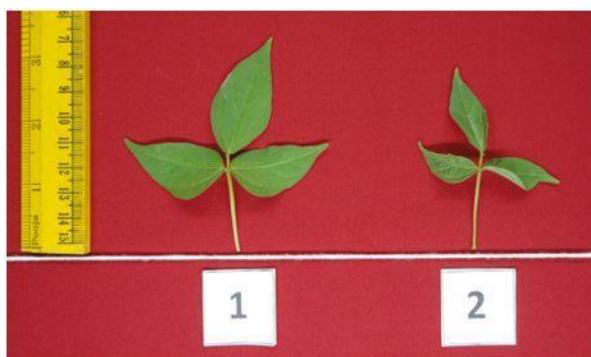
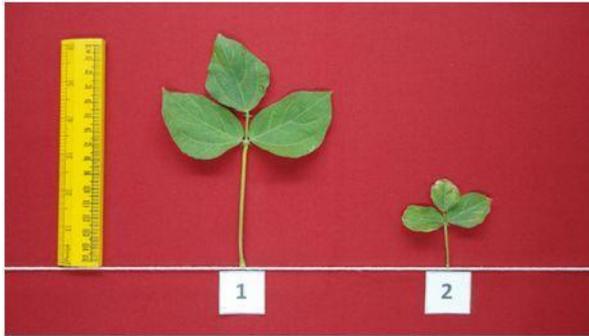


Figure 1: 15 DAS leaves



Figure 2: 30 DAS leaves

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**Figure 3: 45 DAS leaves**



**Figure 4: 60 DAS leaves**



**Figure 5: Chlorosis under UV-B exposure**



**Figure 6: Necrosis under UV-B exposure**



**Figure 7: 45 DAS plant tops**



**Figure 8: Root system and nodules**



**Figure 9: Pods**



**Figure 10: Seeds**

**Plate 1: First fully expanded trifoliate leaves, foliar injuries, plant tops and yield of control (1) and ultraviolet-B (2) irradiated *Vigna mungo* (L.) Hepper var. ADT-3**

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**Table 1: Changes in foliage of *Vigna mungo* (L.) Hepper var. ADT-3 under control and supplementary UV-B exposed conditions**

Treatment	Days after seed sowing	Number of leaves	Total leaf area (cm <sup>2</sup> )	Leaf area index	Specific leaf weight (g <sup>-2</sup> )	Fresh weight of foliage (g)	Dry weight of foliage (g)
Control	15	2	73.60	0.348	0.001	1.729	0.083
	30	3	249.97	0.509	0.006	1.921	0.155
	45	5	644.78	0.735	0.007	1.743	0.501
	60	5	521.20	0.747	0.001	1.464	0.944
UV-B	15	2	29.77	0.411	0.001	1.591	0.255
	30	3	63.83	0.428	0.008	1.424	0.053
	45	4	108.10	0.814	0.001	0.271	0.142
	60	2	157.87	0.383	0.001	0.282	0.225

**Table 2: Changes in growth parameters of *Vigna mungo* (L.) Hepper var. ADT-3 under control and supplementary UV-B exposed conditions**

Treatment	Days after seed sowing	Root length (cm)	Shoot length (cm)	Shoot / root ratio	Root fresh wt. (g)	Shoot fresh wt. (g)	Plant fresh wt. (g)	Root dry wt. (g)	Shoot dry wt. (g)	Plant dry wt. (g)	Relative growth rate
Control	15	7.9	17.5	2.21	1.311	2.115	4.826	0.029	0.136	0.165	-
	30	9.5	21	2.21	1.781	2.614	4.395	0.055	0.289	0.344	0.12
	45	10.5	36	3.42	4.381	4.258	6.639	0.200	1.043	1.243	0.12
	60	12.5	44.5	3.56	7.359	6.343	13.702	0.908	2.376	3.284	0.13
UV-B	15	5.2	16	3.07	2.703	1.824	4.127	0.013	0.091	0.104	-
	30	8.5	18	2.11	1.762	1.911	4.248	0.022	0.122	0.144	0.11
	45	9.5	22.5	2.36	3.234	2.191	5.068	0.078	0.329	0.407	0.11
	60	10	36.1	3.00	4.022	4.022	5.711	0.321	0.625	1.138	0.13

**Table 3: Changes in yield components of *Vigna mungo* (L.) Hepper var. ADT-3 under control and supplementary UV-B exposed conditions**

Treatment	Days after seed sowing	Pod number plant <sup>-1</sup>	Single pod wt. (g)	Pod wt. plant <sup>-1</sup> (g)	Length of the pod (cm)	Seed number pod <sup>-1</sup>	Seed number plant <sup>-1</sup>	Seed mass pod <sup>-1</sup> (g)	Seed mass plant <sup>-1</sup> (g)	Shelling percentage plant <sup>-1</sup>	Harvest index
Control	60	6	0.460	2.192	4.5	7	31	0.221	1.023	25.95	66.76
UV-B	60	3	0.297	0.739	3.5	5	12	0.175	0.390	55.94	54.66

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Supplementary UV-B altered the DNA and protein, which in turn altered the vital metabolisms including photosynthesis reflecting them in the form of reduced yield and nutrition content in the grains (Rajendiran and Ramanujam, 2003; Rajendiran and Ramanujam, 2004).

A technique to ameliorate the UV-B induced suppression in crops needs to be designed in near future as the food security will be in danger if any further increase in ozone depletion occurs in the Earth's atmosphere.

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### **REFERENCES**

- Adamse P and Britz SJ (1992).** Amelioration of UV-B damage under high irradiance I. role of photosynthesis. *Photochemistry and Photobiology* **56** 645 - 650.
- Arulmozhi D and Rajendiran K (2014).** Effect of supplementary ultraviolet-B radiation on nodulation and nitrogen metabolism in *Lablab purpureus* L. var. Goldy. *International Journal of Advanced Biological Research* **4**(3) 343-346.
- Brandle JR, Campbell WF, Sisson WB and Caldwell MM (1977).** Net photosynthesis, electron transport capacity and ultrastructure of *Pisum sativum* L. exposed to ultraviolet-B radiation. *Plant Physiology* **60** 165 - 169.
- Britz SJ and Adamse P (1994).** UV-B induced increase in specific leaf weight of cucumber as a consequence of increased starch content. *Photochemistry and Photobiology* **60** 116 - 119.
- Caldwell MM (1968).** Solar ultraviolet radiation as an ecological factor for alpine plants. *Ecological Monographs* **38** 243 - 267.
- Caldwell MM and Flint SD (1994).** Solar ultraviolet radiation and ozone layer change: Implications for crop plants. In: *Physiology and Determination of Crop Yield*, edited by Boote KJ, Bennett JM, Sinclair TR and Paulson GM, ASA-CSSA-SSSA, Madison WI 487-507.
- Caldwell MM, Bjorn LO, Bornman JF, Flint SD, Kulandaivelu G, Teramura AH and Tevini M (1998).** Effects of increased solar ultraviolet radiation on terrestrial ecosystems. *Photochemistry and Photobiology* **46** 40 - 52.
- Day TA and Demchik SM (1996).** Influence of enhanced UV-B radiation on biomass allocation and pigment concentrations in leaves and reproductive structures of greenhouse - grown *Brassica rapa*. *Vegetatio* **127** 109 - 116.
- Dumpert K and Knacher T (1985).** A comparison of the effects of enhanced UV-B radiation on some crop plants exposed to greenhouse and field conditions. *Biochemie und Physiologie der Pflanzen* **180** 599 - 612.
- Esser G (1980).** Einfluss einer nach Schadstoffimmission vermehrten Einstrahlung von UV-B auf Kulturpflanzen. 2. Versuchsjahr - Bericht Bettelle Institut E. V. Frankfurt, B. F. **63** 984 - 991.
- Francis CA, Flor CA and Prager M (1978).** Effects of bean association on yield component of maize. *Crop Science* **18** 760 - 764.
- Giller YE (1991).** UV-B effects on the development of photosynthetic apparatus, growth and productivity of higher plants. In: *Impact of Global Climatic Changes on Photosynthesis and Plant Productivity*, edited by Abrol YP and Wattal PN 123.
- Huang S, Dai Q, Peng S, Chavez AO, Miranda MLL, Visperas RM and Vergara BS (1997).** Influence of supplementary ultraviolet-B on indole acetic acid and calmodulin in the leaves of rice (*Oryza sativa* L.). *Plant Growth Regulation* **21** 59 - 64.
- Jain VK, Dhingra GK and Ambrish K (1999).** Changes in productivity and biomass partitioning in field grown mungbean with response to supplementary UV-B radiation. In: *Plant Physiology for Sustainable Agriculture*, edited by Srivastava GC, Singh K and Pal M (Pointer Publishers) Jaipur, India 301-308.

### **Research Article**

**Jordan BR (1997).** Effects of UV-B radiation on plants - An overview. In. UV radiation and its Effects: An update, Report of a workshop sponsored by the National Science Strategy Committee for Climate Change. *The Royal Society of New Zealand* **49** 27-28.

**Klaper R, Frankel S and Berenbaum MR (1996).** Anthocyanin content and UV-B sensitivity in *Brassia rapa*. *Photochemistry and Photobiology* **83** 811 - 813.

**Kokilavani V and Rajendiran K (2013).** Ultraviolet-B induced changes in the leaf epidermal and anatomical characteristics of *Vigna mungo* L. var. KM-2. *International Journal of Science and Nature* **5**(1) 126-130.

**Kokilavani V and Rajendiran K (2014a).** Changes in leaf architecture of *Vigna unguiculata* (L.) Walp. cv. BCP-25 after exposure to elevated ultraviolet-B radiation. *International Journal of Science and Nature* **5**(3) 542-546.

**Kokilavani V and Rajendiran K (2014b).** Ultraviolet-B induced changes in the leaf architecture of *Cucumis sativus* L. var. CO 1. *International Journal of Geology, Earth and Environmental Sciences* **4**(2) 208-215.

**Kokilavani V and Rajendiran K (2014c).** Alterations in leaf architecture of *Ocimum sanctum* L. under elevated ultraviolet-B stress. *Global Journal of Bio-Science and Biotechnology* **3**(4) 374-378.

**Kokilavani V and Rajendiran K (2014d).** Influence of elevated ultraviolet-B radiation on the morphology and growth of ten varieties of cowpea. *International Journal of Food, Agriculture and Veterinary Sciences* **4**(3) 87-94.

**Kokilavani V and Rajendiran K (2014e).** Effect of supplementary ultraviolet-B radiation on the yield of ten varieties of cowpea. *International Journal of Geology, Earth and Environmental Sciences* **4**(3) 65-73.

**Kulandaivelu G, Maragatham S and Nedunchezian N (1989).** On the possible control of ultraviolet - B induced response in growth and photosynthetic activities in higher plants. *Physiologia Plantarum* **76** 398 - 404.

**Mark U, Mark SM and Tevini M (1996).** Effects of solar UV-B radiation on growth, flowering and yield of central and southern European maize cultivars (*Zea mays* L.). *Photochemistry and Photobiology* **64** 457 - 463.

**Mark SM and Tevini M (1997).** Effects of solar UV-B radiation on growth, flowering and yield of central and southern European bush bean cultivars (*Phaseolus vulgaris* L.). *Plant Ecology* **128** 114 - 125.

**Mohan P, Bhat MG, Singh NN and Singh P (1992).** Variability of biomass and harvest index in Asiatic (*Gossypium arboreum* L.) and American (*Gossypium hirsutum*) cottons. *Advances in Plant Sciences* **5** 100 - 105.

**Musil CF (1995).** Differential effects of elevated ultraviolet-B radiation on the photochemical and reproductive performances of dicotyledonous and monocotyledonous arid - environment ephemerals. *Plant, Cell & Environment* **18** 844 - 854.

**Nedunchezian N and Kulandaivelu G (1997).** Changes induced by UV-B (280 - 320 nm) radiation in vegetative growth and photosynthetic characteristics in field grown *Vigna unguiculata* L. *Plant Science* **123** 85 - 92.

**Nogues S, Allen DJ, Morison JIL and Baker NR (1998).** UV-B radiation effects on water relations, leaf development and photosynthesis in droughted pea plants. *Plant Physiology* **117** 173 - 181.

**Pearce RB, Brown RK and Blaser RE (1968).** Photosynthesis of *Alfalfa* leaves as influenced by age and environment. *Crop Science* **8** 677 - 680.

**Racey GD, Glerum C and Hutchinson RE (1983).** The practicability of top root ratio in nursery stock characterization. *Forest Chronicle* **59** 240 - 243.

**Rajendiran K (2001).** Amelioration of Ultraviolet-B radiation impacts in green gram by Triadimefon. PhD. Thesis, Pondicherry University.

**Rajendiran K and Ramanujam MP (2000).** Efficacy of triadimefon treatment in ameliorating the UV-B stress in green gram. In: *National Symposium on Environmental Crisis and Security in the New Millennium*, edited by Khan M (National Environmental Science Academy) New Delhi 41-42.

### **Research Article**

- Rajendiran K and Ramanujam MP (2003).** Alleviation of ultraviolet-B radiation-induced growth inhibition of green gram by triadimefon. *Biologia Plantarum* **46** 621-624.
- Rajendiran K and Ramanujam MP (2004).** Improvement of biomass partitioning, flowering and yield by triadimefon in UV-B stressed *Vigna radiata* (L.) Wilczek. *Biologia Plantarum* **48** 145-148.
- Rajendiran K and Ramanujam MP (2006).** Interactive effects of UV-B irradiation and triadimefon on nodulation and nitrogen metabolism in *Vigna radiata* plants. *Biologia Plantarum* **50**(4) 709-712.
- Staxen I and Bornman JF (1994).** A morphological and cytological study of *Petunia hybrida* exposed to UV-B radiation. *Physiologia Plantarum* **91** 735 - 740.
- Sudaroli Sudha J and Rajendiran K (2013a).** Effect of elevated UV-B irradiation on the nodulation and nitrogen metabolism in *Sesbania grandiflora* (L.) Pers. *International Journal of Science and Nature* **4**(4) 664 - 667.
- Sudaroli Sudha J and Rajendiran K (2013b).** Effect of elevated UV-B irradiation on the nodulation and nitrogen metabolism in *Vigna unguiculata* (L.) Walp. c.v. BCP-25. *International Journal of Food, Agriculture and Veterinary Sciences* **3**(3) 77-81.
- Sullivan JH, Teramura AH and Dillenburg LR (1994).** Growth and photosynthetic responses of field-grown sweetgum (*Liquidambar styraciflua*) seedlings to UV-B radiation. *American Journal of Botany* **81** 826 - 832.
- Tevini M and Teramura AH (1989).** UV-B effects on terrestrial plants. *Photochemistry and Photobiology* **50** 479 - 487.
- Tevini M, Mark U, Fieser G and Mark SM (1991).** Effects of enhanced solar UV-B radiation on growth and function of selected crop plant seedlings. In: *Photobiology*, edited by Riklis E (Plenum Press) New York 635-649.
- Vijayalakshmi R and Rajendiran K (2014).** Impact of ultraviolet-B radiation on nodulation and nitrogen metabolism in *Phaseolus vulgaris* L. cv. Prevail. *International Journal of Advanced Biological Research* **4**(3) 339 - 342.
- Williams RF (1946).** The physiology of plant growth with special reference to the concept of net assimilation rate. *Annals of Botany* **10** 41 - 42.
- Ziska LH, Teramura AH and Sullivan JH (1992).** Physiological sensitivity of plants along an elevational gradient to UV-B radiation. *American Journal of Botany* **79** 863 – 871.