# AN INVESTIGATION OF THE PHYTOCHEMICAL AND ELEMENTAL CONTENT OF STEM BARK OF *VITEX DONIANA* SWEET (BLACK PLUM)

Mustapha A. Tijjani<sup>1</sup>, \*Fanna I. Abdurahaman<sup>1</sup>, Irfan Z. Khan<sup>1</sup> and Umar K. Sandabe<sup>2</sup>

<sup>1</sup>Department of Chemistry, Faculty of Science, University of Maiduguri P.M.B 1069, Maiduguri, Borno State, Nigeria

<sup>2</sup>Department of Veterinary Physiology, Pharmacology and Biochemistry, University of Maiduguri P.M.B 1069, Maiduguri, Borno State, Nigeria \*Author for Correspondence

### ABSTRACT

Phytochemical and elemental contents of stem bark of V. doniana were investigated. The stem bark of V.doniana (5kg) was macerated with ethanol and thereafter sequentially extracted using chloroform, ethyl acetate and n-butanol. Moisture content and ash content were determined. Phytochemical analysis of the air dried matter, ethanol, chloroform, ethyl acetate and n-butanol of the stem bark extracts of V. doniana were carried out using standard procedures. The stem bark of V. doniana was ashed, digested and analysed for heavy and trace elemental content using atomic absorption and flame emission spectroscopy (AAS and EES). The percentage moisture content and ash value were 7% and 3%. The extracts concentrate yields of ethanol, chloroform, ethylacetate and n-butanol were 37% w/w, 1.25% w/w, 2.4% w/w, 3.36% w/w. Preliminary phytochemical screening revealed the presence of tannins, saponins, steroid, terpenes, phloba-tannins, carbohydrate, flavonoids and cardioative-glycosides in the dry matter, ethanolic, ethylacetate and n-butanol fractions. The chloroform fraction has only cardioactive glycosides and carbohydrate. Alkaloids and anthraquinone were absent in all the extracts. Results of elemental contents determination indicates that the concentration in ug/g of some trace essential elements such as Mangnese (Mn), Magnesium (Mg), Potassium (K) and Calcium (Ca) in the stem bark of vitex doniana to be within safety limits reported by World Health Organization. However the concentration of Potasium (K) is slightly above the WHO recommended level. Non-essential and toxic metals such as lead (Pb) and cadmium (Cd) were not detected while Iron (Fe), Zinc (Zn), Cobalt (Co), Sodium (Na), Copper (Cu) and Phosphrus (P) are found in low concentration. These essential elements detected in the stem bark of V. doniana are very much required by our bodies for biological and physiological functions that are necessary for maintenance of good health while the phytochemicals reported are implicated in having many pharmacological properties. Further research is going on purification, characterization, structural elucidation and assaying some of the pharmacological activities.

Key Words: Phytochemical, Vitex doniana, Elemental

## **INTRODUCTION**

Nearly all cultures of the world, both ancient and the recent have heavily depended on plants as a therapeutic agents used in various forms. Plants play a major role in the treatment of diseases and still remain the foremost alternative for a large majority of people. (Adiaratou et al., 2005). The knowledge of plants if wisely utilized could draw out promising herbal leads. The World Health Organization (WHO, 1996) has reported that about 80% of the world population relies on traditional medicine to cure ailments (Farnsworth et al., 1985 and Harsha et al., 2002). Medicinal plants is defined as any plant in which one or more of its organs contain substances that can be used for therapeutic purpose and which are precursors for the synthesis of new drug (Sofowora,1993). Phytochemicals are rich source of organic substances which are used for production of many drugs. Investigation into therapeutic efficacy of plants is therefore urgently needed because of widely usage, acceptability and so many claimed activities for treatment of ailments and diseases. The phytochemicals that have medicinal importance are the secondary metabolites.

These secondary metabolites includes phenolic compounds such as tannins and flavonoids, glycosides such as cardiac, flavonoids, steroidal and saponins. Terpenes, alkaloids, volatile oil, gums and mucilage. These organic compounds were extracted, isolated, purified and subjected to detailed investigation for their medicinal/pharmacological properties by scientist using standard procedures so as to prove the claim therapeutic properties. Elemental research has definitely been part of this explosion of scientific knowledge (Said et al., 1996). Several studies have reported elemental contents in plant extracts which are consumed as herbal health drink or in orthodox medicine (Rajua et al., 2006). The qualitative estimation of various trace element concentration is important for determining the effectiveness of medicinal plants in treating various diseases and also to understand the pharmacological action (Rajua et al., 2006). Trace elements that are necessary for human health include Iron, iodine, copper, manganese, zinc, molybdenum, selenium and chromium etc (Hendler and Shelder, 1990). Herbs are not only provide us with chemicals of medicinal value but also provide us nutritional and trace elements (Zafar et al., 2010.)

*Vitex doniana* Sweet, a plant commonly known black plum, in English, Prunier noir in French, dinya in Hausa, ucha koro in Igbo, oori-nla in yoruba and ngarmi in Kanuri is a medium-sized deciduous tree, 8-18 m high, with a heavy rounded crown and a clear bole up to 5 m. V. *doniana* is from *Verbenaceae* family and abundantly occurring in savannah regions. It is a deciduous forest tree of coastal woodland, riverine and lowland extending as high as upland grassland, secondary forests, and dry forests. It can be found throughout tropical Africa. (Han-Jurgen, 1990; Hutchinson and Dalziel, 1964; Keay, 1964 and Kokwaro, 1976). Studies have reported the elemental and phytochemistry of root bark and leaves of V. *doniana*, however there is a scanty report on the stem bark. The objective of this research is to investigate the ash, moisture, phytochemical and elemental contents of stem bark of V. *doniana*.

## MATERIALS AND METHODS

## Sample collection and identification

The stem bark of Vitex doniana leaves were collected in Kawuri village of Konduga (11°39'6"N 13°25'10"E) Local Government Area of Borno state of Nigeria. The plant specimen was identified and authenticated by a Plant Taxonomist, Prof. S.S. Sanusi of the Department of Biological Sciences, Faculty of Science, University of Maiduguri. The herbarium specimen with a voucher number 555C was deposited at the Post Graduate Research Laboratory, Department of Chemistry. The stem bark of the Vitex doniana was cleaned and air-dried in the laboratory. Five kilogram (5kg) of the stem bark of Vitex doniana was pulverised into a coarse powder using mortar and pestle.

## Estimation of moisture content

Five grammes (5g) of V. *doniana* stem bark were placed in a weighed watch glass and dried in the oven at 80°C to constant weight using standard procedures with constant checking for 2 hours. The percentage moisture content was then determined (AOAC,1984 and Radojevic and Baskin, 1999).

## Ash value determination

Five grammes (5g) of V. *doniana* stem bark were put in a weighted acid washed and air dried porcelain crucible in a hot spot furnace, and the temperature was raised slowly until it reached 500°C. it was then left there for about 4 hours. The crucible was removed from the furnace after ashing with a pair of thongs and weighted. The percentage ash value was then determined. (AOAC,1984, Radojevic and Baskin, 1999).

#### Extraction of stem bark

The weighed powdered air-dried sample (5kg) was macerated with ethanol for five days and partitioned sequentially with choroloform, ethyl acetate, and n-butanol. The extract obtained was concentrated to dryness in vacuo at  $40^{\circ}$ C using a rotary evaporator. The extract concentrate was labeled and the percentage yields were calculated in w/w.

#### Phytochemical analysis

The air died matter, crude extracts of the choroloform, ethyl acetate, n-butanol and ethanol were subjected to qualitative chemical screening for identification of the primary and secondary metabolites such as flavonoids, alkaloids, sterols, triterpenes, saponins, anthracenosides, tannins, polyuronides, emodol, etc as described by (Ioan, 1982; Sofowora, 1993 and Trease and Evans 2002).

#### Elemental Contents Evaluation

The air dried sample (10g) was dried to a constant weigh in an oven at 80°C. The sample (5g) was then ashed in a hotspot furnace at 500°C for 3 hours. The ashed material was then digested using standard procedures (Radojevie and Baskin 1999) and analysed for the determination of trace and heavy elements present in *Vitex doniana* stem bark using a combination of flame emission spectrometry (FES) (GallenKamp) and Atomic Absorption Spectrometry (AAS) (SPG Unicam Model No.1) at appropriate wavelength, temperature and lamp current for each element under study (Radojevie and Baskin 1999). Flame emission spectrometry (FES) (Gallen Kamp FGA 330) was use to determine sodium and potassium. Elements such as magnesium, calcium, iron, lead zinc, manganese, cadmium and copper was determined by atomic absorption spectrometry (AAS).

## **RESULTS AND DISCUSSION**

## Moisture Content and Ash Value

The percentage moisture content and ash value are 7% and 3%. The total ash could serve as a standard in determining the amount of residual substances not volatilized when the drug is ignited (African Pharmacopoeia, 1986).

The extracts concentrate yields of ethanol, chloroform, ethylacetate and n-butanol were 37 %w/w, 1.25% w/w, 2.4% w/w and 3.70%. As seen from table 2 and 3, the results of the pre-liminary phytochemical analysis revealed the presence of tannins, phlobatannins, saponins, carbohydrates, cardioactive glycoside, flavonoids, steroids and terpene in the dry matter, ethanolic, ethylacetate and nbutanol fractions. The chloroform fraction has only cardioactive glycosides and carbohydrates. Alkaloids and anthracenosides were conspicuously absent in all the extracts. Plants are rich bio resources of drugs. A number of interesting outcomes have been found with the use of a mixture of -natural products or plant extracts to treat diseases (Gibbons, 2003). Phytochemical studies are important aspects of medicinal plants research and are used in evaluating the presence of the chemical constituents in plants (Abdurahaman, 2004). The phytochemicals that have medicinal importance are the secondary metabolites. These secondary metabolites includes phenolic compounds such as tannins and flavonoids, glycosides, cardioactive glycoside, steroidal glycosides and saponins. Terpenes, alkaloids, volatile oil etc. Tannin have astringent properties which are important in wound healing (Pondromoli and Grazi, 1969). Flavonoid compounds are is implicated in having an antipyretic property, anti-inflammatory and antioxidant property. (Trease and Evans, 2002). Saponins have an expectorant action that is very useful in management of upper respiratory tract. Cardioactive glycosides are naturally cardioactive drugs used in treatment of congestive heart failure and cardiac arrhythmia (Brian et al., 1985)

Table 1. Percentage yield, colour and texture of EtOH	, CHCl <sub>3</sub> , EtOAc and n-butanol
extracts of stem bark of V. doniana	

Extract fraction	%yield(w/w)	Colour	Texture
EtOH	37%	black	gummy
CHCl <sub>3</sub>	1.25%	dark brown	gummy
EtOAc	2.40%	light brown	gummy
n-butanol	3.70%	dark brown	powder

## Table 2. Phytochemistry of the dry matter

S/N	Class of Chemical Components	Results		
1.	Test of Alkaloids			
	Dragendoff reagent	-		
	Mayers reagent	-		
2.	Test for Flavonoid			
	Shinoda Test	-		
	Lead acetate test	++		
	Sodium hydroxide test	++		
	Ferric Chlorides test	+		
3.	Test for carbohydrate			
a.	General Test (Molish test)	+		
b.	Test of monosaccharide	-		
c.	Test for reducing sugar (Fehling test)	++		
d.	Combine reducing sugar test	++		
e.	Test for ketoses	+		
f.	Test for pentose	+		
4.	Test for tannins			
	Ferric chloride	+		
	Lead acetate	+		
	Hydrochloric acid test	-		
5.	Test for free Anthraquinones (Bontrase)	-		
	Test for combined anthraquinone	-		
6.	Test for cardio active glycoside			
	Salkowski test	+		
	Liebermann Burchard test	+		
7.	Terpenoid test	+		
8.	Test for soluble starch	+		
9.	Test for phlobatannins	-		
10. Test for saponins				
i.	Frothing test	+		
ii. <u>Fehling test</u> ++				
Ke	y - Absent			

+ Present in low concentration

++ Present in moderate concentration

Table 3. Phytochemistry of ethanolic (EtOH), chloroform (CHCl<sub>3</sub>) ethylacetate (EtOAc), and n-butanol extracts.

ITest of Alkaloids Dragendoff reagentMayersreagent2.Test for Flavonoid Shinoda Test+++Lead acetate test++-+++Solium hydroxide test++-+++Solium hydroxide test++-+++Solium hydroxide test++-+++Solium hydroxide test++-+++Solium hydroxide test++-+++Solium hydroxide test++-+++General Test (Molish test)+-+++Test for carbohydrate+Combine reducing sugar (Fehing test)++++Test for tannins+++Ferric chloride++-++++++Hydrochloric acid test5.Test for cardio-active glycoside Salkowski test++++++++++++++++++++++++++++++++++++++ </th <th>S/N</th> <th>Class of Chemical Components</th> <th>EtOH</th> <th>CH</th> <th>ICl<sub>3</sub></th> <th>EtOAc</th> <th>n- butanol</th>	S/N	Class of Chemical Components	EtOH	CH	ICl <sub>3</sub>	EtOAc	n- butanol
Dragendoff reagent       -       -       -       -       -       -         Mayers reagent       -       -       -       -       -       -         2. Test for Flavonoid       Shinoda Test       -       +       +       +       +         Shinoda Test       +       +       +       +       +       +       +         Solum hydroxide test       ++       -       +       +       +       +       +         Solum hydroxide test       ++       -       + <th>1.</th> <th>Test of Alkaloids</th> <th></th> <th></th> <th></th> <th></th> <th></th>	1.	Test of Alkaloids					
Mayers reagent2. Test for FlavonoidShinoda Test++Shinoda Test++Lead acetate test++-+++Sodium hydroxide test++-+++Ferric Chlorides test++-+++Test for carbohydrate+++General Test (Molish test)+-+++Test for raducing sugar (Fehling test)+++Test for reducing sugar (Fehling test)+++Test for ketoses+++Test for tannins++Ferric chloride++-+++++Lead acetate++-+++Hydrochloric acid test5. Test for free Anthraquinones (Bontrase)6. Test for cardio-active glycosideSalkowski test++++++++++++++1. Test for soluble starch+++++++++9. Test for soluble starch++++++10. Test for splobatannins+-+++++Frobing test++-++++++11. T		Dragendoff reagent		-	-	-	-
2. Test for Flavonoid Shinoda Test + + + Lead acetate test ++ - ++ Sodium hydroxide test ++ - ++ + Ferric Chlorides test ++ - ++ ++ Ferric Chlorides test ++ - ++ ++ 3. Test for carbohydrate General Test (Molish test) ++ - + ++ ++ Test of monosaccharide(Barfoed test) ++ ++ Test of monosaccharide(Barfoed test) ++ - + - ++ ++ Combine reducing sugar (Fehling test) ++ + ++ ++ Test for ketoses ++ ++ ++ Test for ketoses ++ ++ ++ Test for pentose ++ ++ ++ Hydrochloric acid test ++ - ++ ++ ++ 5. Test for free Anthraquinones (Bontrase) 6. Test for free Anthraquinones (Bontrase) 7. Test for free Anthraquinones (Bontrase) ++ ++ ++ ++ ++ Liebermann Burchard test ++ ++ ++ ++ ++ 8. Teerpenoid test ++ ++ ++ ++ ++ 9. Test for soluble starch ++ ++ ++ ++ ++ 10. Test for soluble starch ++ ++ ++ ++ ++ Frothing test ++ - ++ ++ ++ ++ Frothing test ++ - ++ ++ ++ ++ ++ Frothing test ++ - ++ ++ ++ ++ ++ ++ ++ ++ ++ ++ ++ +		Mayers reagent		-	-	-	-
Shinoda Test++Lead acetate test++-+++Sodium hydroxide test++-++++Ferric Chlorides test++-++++ <b>3. Test for carbohydrate</b> -++++General Test (Molish test)+-+++Test for monosaccharide(Barfoed test)Test for reducing sugar (Fehling test)++++Test for reducing sugar (Fehling test)++++Test for ketoses+++Test for ketoses++Test for tannins+Ferric chloride++-++++Hydrochloric acid test5. Test for free Anthraquinones (Bontrase)6. Test for cardio-active glycosideSalkowski test++++++++++Liebermann Burchard test++++++++8. Teerpenoid test+++++++9. Test for soluble starch++++1. Test for splobatannins+-++1. Test for soluble starch+++++1. Test for soluble starch+++++1. Test for soluble starch++++++1. Test for soluble	2.	Test for Flavonoid					
Lead acetate test++-++++Sodium hydroxide test++-+++Ferric Chlorides test++-+++++3. Test for carbohydrateGeneral Test (Molish test)+-+++Test for monosaccharide (Barfoed test)Test for reducing sugar (Fehling test)++++Test for reducing sugar (Fehling test)++++Test for ketoses++++Test for ketoses++Test for tannins+Ferric chloride++-++++Lead acetate++f. Test for free Anthraquinones (Bontrase)f. Test for cardio-active glycosideSalkowski test++++++++Liebermann Burchard test++++++8. Terpenoid test+++++9. Test for soluble starch++++10. Test for phlobatannins+-++11. Test for saponins+-++++Frothing test++-++++Hehling test++-++++Test for saponins++-++++Test for saponins++-++++Test fo		Shinoda Test		-	-	+	+
Sodium hydroxide test++-++Ferric Chlorides test++-+++++ <b>3. Test for carbohydrate</b> General Test (Molish test)+-+++Test of monosaccharide(Barfoed test)Test for reducing sugar (Fehling test)++++Combine reducing sugar test++++Test for reducing sugar test++++Test for katoses++Test for tannins+Ferric chloride++-++++Hydrochloric acid test5. Test for free Anthraquinones (Bontrase)6. Test for cardio-active glycoside Salkowski test++++++++8. Terpenoid test++++++++9. Test for soluble starch+++++++10. Test for soluble starch+++++11. Test for saponins+-+++++Frothing test++-+++++		Lead acetate test		++	-	++	++
Ferric Chlorides test++-+++++3. Test for carbohydrate General Test ( Molish test)+-+++Test of monosaccharide( Barfoed test)Test for reducing sugar (Fehling test)++++Combine reducing sugar test++++Test for ketoses+++Test for ketoses++Test for pentose+4. Test for tannins+Ferric chloride++-++++Hydrochloric acid test5. Test for free Anthraquinones (Bontrase)6. Test for cardio-active glycoside Salkowski test++++++++8. Terpenoid test++++++++9. Test for soluble starch++++10. Test for saponins+-+++11. Test for saponins+-+++++Frothing test++-+++++		Sodium hydroxide test		++	-	+	+
3. Test for carbohydrate General Test (Molish test) + - + + + + + + + + + + + + + + + + +		Ferric Chlorides test		++	-	+++	++
General Test (Molish test)+-+++Test of monosaccharide( Barfoed test)Test for reducing sugar (Fehling test)++++Combine reducing sugar test++++Test for ketoses+++Test for ketoses++Test for ketoses++Test for tannins+Ferric chloride++-++++Lead acetate++-++++Hydrochloric acid test5.Test for free Anthraquinones (Bontrase)6.Test for cardio-active glycosideSalkowski test++++++++++Liebermann Burchard test++++++++9.Test for soluble starch++++++9.Test for phlobatannins+-+++10.Test for soluble starch++++11.Test for solubine starch++-++++11.Test for solubine test++-++++Frohing test++-++++++Frohing test++-++++++Hydrochloric acid test++-++++	3.	Test for carbohydrate					
Test of monosaccharide( Barfoed test)Test for reducing sugar (Fehling test)++++Combine reducing sugar test++++Test for ketoses++Test for ketoses++Test for pentose+4. Test for tannins+Ferric chloride++-++++Lead acetate++-++++Hydrochloric acid test5. Test for free Anthraquinones (Bontrase)6. Test for cardio-active glycosideSalkowski test++++++++Liebermann Burchard test++++++8. Terpenoid test++++++9. Test for soluble starch++++10. Test for phlobatannins+-+11. Test for saponins+Frothing test++-+++Fehling test++-++		General Test (Molish test)		+	-	+	++
Test for reducing sugar (Fehling test) $++$ $  ++$ Combine reducing sugar test $++$ $  ++$ Test for ketoses $+$ $  +$ Test for pentose $   +$ 4. Test for tannins $   +$ Ferric chloride $++$ $ ++$ $++$ Lead acetate $++$ $ ++$ $++$ Hydrochloric acid test $   -$ 5. Test for free Anthraquinones (Bontrase) $   -$ 6. Test for cardio-active glycoside $   -$ 7. Test for cardio-active glycoside $++$ $++$ $++$ $++$ Salkowski test $++$ $++$ $++$ $++$ 8. Terpenoid test $++$ $++$ $++$ $++$ 9. Test for soluble starch $+$ $+$ $+$ $+$ 10. Test for saponins $+$ $ +$ $+$ Frothing test $++$ $ ++$ Frothing test $++$ $-$ Frothing test $++$ $ ++$ Frothing test </td <td></td> <td>Test of monosaccharide( Barfoed test)</td> <td>)</td> <td>-</td> <td>-</td> <td>-</td> <td>-</td>		Test of monosaccharide( Barfoed test)	)	-	-	-	-
Combine reducing sugar test $++$ $  ++$ Test for ketoses $+$ $  +$ Test for pentose $   +$ 4. Test for tanninsFerric chloride $++$ $ ++$ Ferric chloride $++$ $ ++$ $++$ Lead acetate $++$ $ ++$ $++$ Hydrochloric acid test $   -$ 5. Test for free Anthraquinones (Bontrase) $   -$ 6. Test for cardio-active glycoside $   -$ 7. Test for cardio-active glycoside $++$ $++$ $++$ Salkowski test $++$ $++$ $++$ 8. Terpenoid test $++$ $++$ $++$ 9. Test for soluble starch $+$ $+$ $++$ 10. Test for phlobatannins $+$ $ +$ 11. Test for saponins $++$ $ ++$ Frothing test $++$ $ ++$ $+$ $++$ $ ++$		Test for reducing sugar (Fehling test)		++	-	-	++
Test for ketoses++Test for pentose+4. Test for tannins+Ferric chloride++-++++Lead acetate++-++++Hydrochloric acid test5. Test for free Anthraquinones (Bontrase)6. Test for cardio-active glycoside7. Test for cardio-active glycosideSalkowski test++++++Liebermann Burchard test+++++9. Test for soluble starch+++10. Test for phlobatannins+-+Frothing test++-++++Frothing test++-++++-++++		Combine reducing sugar test		++	-	-	++
Test for pentose+4. Test for tanninsFerric chloride++-++++Ferric chloride++-++++Lead acetate++-++++Hydrochloric acid test5. Test for free Anthraquinones (Bontrase)6. Test for combine anthraquinone7. Test for cardio-active glycosideSalkowski test++++++++Liebermann Burchard test++++++9. Test for soluble starch++++10. Test for phlobatannins+-+H-++++++Frothing test++-+++Fehling test++-+++++++++		Test for ketoses		+	-	-	+
4. Test for tannins Ferric chloride $+++$ - $++$ $++$ Lead acetate $+++$ - $++$ Hydrochloric acid test 5. Test for free Anthraquinones (Bontrase) 6. Test for cardio-active glycoside Salkowski test $++$ $++$ $++$ $+$ Liebermann Burchard test $++$ $++$ $++$ $+$ 8. Terpenoid test $++$ $++$ $++$ $++$ 9. Test for soluble starch $+$ $+$ $++$ $++$ $++$ 10. Test for phlobatannins $+$ $ +$ $+$ $++$ 11. Test for saponins Frothing test $++$ $++$ $ ++$ $++$		Test for pentose		-	-	-	+
Ferric chloride++-++++Lead acetate++-++++Hydrochloric acid test5. Test for free Anthraquinones (Bontrase)6. Test for combine anthraquinone7. Test for cardio-active glycosideSalkowski test+++++++Liebermann Burchard test+++++8. Terpenoid test+++++9. Test for soluble starch+++10. Test for phlobatannins+-+11. Test for saponins++-++Frothing test++-++++++-++	4.	Test for tannins					
Lead acetate Hydrochloric acid test++ +  + + + +++ + + +5. Test for free Anthraquinones (Bontrase) 6. Test for combine anthraquinone- -  -  - -6. Test for cardio-active glycoside Salkowski test Liebermann Burchard test++ ++++ ++++ +8. Terpenoid test 9. Test for soluble starch 11. Test for phlobatannins H++ + +++ +++ + +10. Test for saponins Frothing test H++ + +- +++ + +		Ferric chloride		++	-	++	++
Hydrochloric acid test5. Test for free Anthraquinones (Bontrase)6. Test for combine anthraquinone7. Test for cardio-active glycosideSalkowski test++++++++++Liebermann Burchard test++++++++8. Terpenoid test+++++++9. Test for soluble starch+++++10. Test for phlobatannins+-++11. Test for saponins++-++++Frothing test++-++++		Lead acetate		++	-	++	++
5. Test for free Anthraquinones (Bontrase)6. Test for combine anthraquinone7. Test for cardio-active glycoside Salkowski test++++++++++Liebermann Burchard test++++++++++8. Terpenoid test++++++++9. Test for soluble starch++++++10. Test for phlobatannins+-++11. Test for saponins+++-++++Frothing test+++-++++		Hydrochloric acid test		-	-	-	-
<ul> <li>7. Test for cardio-active glycoside Salkowski test</li> <li>8. Terpenoid test</li> <li>9. Test for soluble starch</li> <li>9. Test for phlobatannins</li> <li>9. Test for saponins</li> <li>9. Frothing test</li> <li>9. Frothing test</li> <li>9. Hert for saponins</li> <li>9. Frothing test</li> <li>9. Hert for saponins</li> <li>9</li></ul>	5	Test for free Anthroquinones (Bontrose)					
<ul> <li>7. Test for cardio-active glycoside Salkowski test ++ ++ ++ ++ ++</li> <li>8. Terpenoid test ++ ++ ++ ++</li> <li>9. Test for soluble starch ++ ++ ++</li> <li>9. Test for phlobatannins ++ ++ ++</li> <li>10. Test for phlobatannins ++ ++ ++</li> <li>11. Test for saponins</li> <li>Frothing test +++ - ++ ++</li> </ul>	5. 6	Test for combine anthraquinone		_	_	_	
7. Test for cardio-active glycoside Salkowski test++++++Salkowski test++++++Liebermann Burchard test++++++8. Terpenoid test+++++9. Test for soluble starch++++10. Test for phlobatannins+-+11. Test for saponins++-++Frothing test++-++++-++++++++	0.	Test for complex antil aquillone		-	-	_	-
Salkowski test $++$ $++$ $++$ $++$ $++$ Liebermann Burchard test $++$ $++$ $+$ $+$ 8. Terpenoid test $+$ $++$ $++$ $++$ 9. Test for soluble starch $+$ $+$ $+$ $+$ 10. Test for phlobatannins $+$ $ +$ $+$ 11. Test for saponins $+$ $ +$ $+$ Frothing test $++$ $ +$ $++$ $ +$ $+$ $+$	7.	Test for cardio-active glycoside					
Liebermann Burchard test $++$ $++$ $+$ $+$ 8. Terpenoid test $+$ $++$ $++$ $++$ 9. Test for soluble starch $+$ $+$ $+$ $+$ 10. Test for phlobatannins $+$ $ +$ $+$ 11. Test for saponins $+$ $ +$ $+$ Frothing test $++$ $ ++$ Frothing test $++$ $ +$ ++ $ +$		Salkowski test		++	++	+	++
8. Terpenoid test+++++++9. Test for soluble starch++++10. Test for phlobatannins+-++11. Test for saponins+-+++Frothing test++-++++Frothing test++-++++		Liebermann Burchard test		++	++	+	+
9. Test for soluble starch++++10. Test for phlobatannins+-++11. Test for saponins++++Frothing test++-++++Fehling test++-+++	8.	Terpenoid test		+	++	++	++
10. Test for phlobatannins+-++11. Test for saponins++-++++Frothing test++-++++Fehling test++-+++	9.	Test for soluble starch		+	+	+	+
11. Test for saponins         Frothing test       ++       -       ++       ++         Fehling test       ++       -       +       ++	10.	Test for phlobatannins		+	-	+	+
Frothing test++-++++Fehling test++-+++	11.	Test for saponins					
Fehling test ++ - + ++		Frothing test		++	-	++	++
		Fehling test		++	-	+	++

Key - Absent

+ *Present in low concentration* 

++ Present in moderate concentration

The elemental analysis results in figure 1 and appendix I indicates that the concentration of some trace essential elements such as Mn, Mg, K and Ca in the Stem bark of *vitex doniana* appeared to be within safety limits reported by World Health Organization (1996). However the concentration of K is slightly **Appendix 1**. Concentrations of trace and heavy elements in digested sample of stem bark of *Vitex doniana* 

ELEMENTS	Concentration (ug/g)	WHO Standard (1999) (ug/g)
Potasium (K	121	10-100
Sodium (Na)	163	400-500
Mangnesium (Mg)	103	100-200
Iron (Fe)	8.4	50-5000
Zinc (Zn)	6.0	150-20000
Copper (Cu)	13.8	100-300
Calcium (Ca)	5423	3600-80000
Suphur (S)	63	8-500
Cobalt (Co)	7.8	100
Lead (Pb)	ND	5-30
Phosphorus (P)	22	100-200
Mangnese (Mn)	560	100-20000

above the WHO recommended level. Trace elements are chemical elements required by our bodies for biological and physiological functions that are necessary for maintenance of good health. The elements



present in the plant may be an indication of the types of minerals presents in the soil. Mineral elements are essential in many vital processes in both plants and animals. According to Kaneez et al., (2003), elements such as Mg in plants lowers the cholesterol level and alleviate heart diseases, Fe deficiency is associated with myocardial infection while Zn is important in wound healing. Mn is essential for normal functioning of central nervous system and is good anti-oxidant (Bibi et al., 2006). Ca is needed in the development of bone teeth, regulates heart rhythm, helps in blood clotting, maintain proper nerve and muscle functions and lowers the blood pressure (Badahur et al., 2011). Non-essential and toxic metals

such as Pb and Cd were not detected in the stem bark *Vitex doniana*. Fe, Zn, Co, Na, Cu and P are found in low concentration. Further research are going on in purification, characterization and structural elucidation of active component in the stem bark of V. *doniana* and some pharmacological studies.

## ACKNOWLEDGMENTS

We wish to acknowledge the technical assistance of Mr Fine Akawo of the Research Laboratory of Department of Chemistry, University of Maiduguri and Mr. Samuel Chabiri of National Agency for Food and Drug, Administration and Control, Maiduguri Laboratory.

## REFERENCES

Abdurahaman, FI (2004). Studies on the chemical contents and Pharmacological activities of the root bark of *Vitex doniana* Sweet (Black Plum) *PhD. Thesis.* pp.33-39 University of Maiduguri.

Adiaratou T, Drissa D, Seydou D, Hilde B, and Berit SP (2005). Ethnopharmacological survey of different uses of seven medicinal plants from Mali, (West Africa) in the regions *Doila, Kolokani* and Silby. *Journal of Ethnobiology and Ethnomedicine* 1(7).

**AOAC**, (1984). Official method of Analysis (15<sup>th</sup> edn). Association of Official Analytical Chemist. Washington, DC USA.

African Pharmacopoeia, (1986). OAU/STRC, Publication No.2 1: 86-88

Bahdhahur, A, Chaudry Z, Guljam M, Danish M, Rahaman A, Rafiq A, Khan A, Shah K, Ullah I., Zahr S, Alli. F, Tahra and Guja F(2001). *African Journal of Pharmacy and Pharmacology* **5**(8) pp.1157-1161.

**Bibi S, Dastagir G, Hussain F and Sanuallah P (2006).** Elemental composite of *Violaodaranta Linn. Park Journal of Plant Science* **12** (2) pp.141-143

Brian, F H, Thomas-Bigger, J and Goodman, G (1985). The Pharmacological Basis of Therapeutics, Macmillan, 7th edition New York, NY, USA. 123-124.

Farnsworth, N R, Akerele, O, Bingle, A S, Soejarto, D D and Guo Z (1985). Medicinal plants in therapy. *Bulletin of the World Health Organization (WHO)* 63(6) pp.965-981.

**Gibbons S** (2003). An overview of plant extracts as potential therapeutics. *Expert Opinion on Therapeutic Patents*, **13**(4) pp.489–497.

**Hans-Jurgen Von Maybell (1990).** Trees and Shrubs of the Sahel. Their characteristics and uses. Verlag Josef Magerat Scientific Book. Muhestrable 9, n-6992 Weikershhein, pp.397.

Harsha V H, Hebbar S S, Hegde G R , and Shripathi V (2002). Ethnomedical knowledge of plants used by Kunabi Tribe of Karnataka in India. *Fitoterapia* 73(4) pp.281-287.

Hendler E and Sheldon S (1990). The Doctors vitamin and mineral. Enclyclopedia. New York NY. *Simon and Schuster*, pp.112-107

Hutchinson, J and Daziel, J M (1964). Flora of West Tropical Africa. Grown Agents. London pp.13

**Ioan, I C (1982).** Methodology for Analysis of Vegetables drugs. Chemical Industries Branch Division of Industrial Operations. UNIDO, Romania 128-141.

Kaneez F A, Quadirrun M, Kalharo M A, Khala and Badar D (2003). Determination of major trace elements in *Artemisia elegantissima* and *Rhazastiulca* and its use. *Park Journal of Plant Science and Industrial Research* **45** pp.291-293

Keay R W J, Onochie C F A and Stanfied D F (1964). Nigerian Trees. (Federal Government Printers. Lagos) 1. pp. 124.

Kokwaro J O. (1976). Medicinal plants of East Africa. East African Literature Bureau. Kenya pp.98-111 Radojeviv M and Baskin V N (1999). Practical Environmental Analysis 1<sup>st</sup> ed. *The Royal Society of Chemistry* (Cambridge, U.K) pp.378-408.

Rajua G JN, Saritha P, Murtya G A V R, Kumar M R, Reddya B S, Charlesa M J Lakshiminayayana S., Reddya SB, and Vijaybbu V (2006). Estimation of trace elements in some anti-diabetic medicinal plants using PIXE Technique. *Applied Radiation and Isotopes* 64 pp.893-900

Saids S, Saeed H M A, Silva L A Zubairy H N and Ban Z (1996). Medicinal Herbal Dugs. A text book for Medical Student and Doctors. Published by Hamdard foundation Pakistan. 74600, Pakistan 1; pp.272-291.

**Sofowora A (2000).** Medicinal Plant and Traditional Medicine in Africa. *Spectrum edition Books* Ltd 3<sup>RD</sup> edn, Ibadan26-100 and 289

**Pontrimoli, A and Grazi E (1969).** A method of assaying liver Hexose monophosphate Oxidation. *Journal of Comprehensive Biochemistry* pp.33-26

**Trease, GE and Evans WC (2002).** Pharmacognosy. 15<sup>th</sup> edn. *Saunders publishers*, London.pp.34-37 **WHO (1996).** Guidelines for elemental concentration. Trace elements in health and human Nutrition pp. 50-228.

Zafar M, Khan M A, Ahmad M, Jan G, Sultana S, Ullah K, Marwat S K Ahmad F, Jabeen A, Nazir A, Abbasi A M, and Ullah Z (2010). Elemental analysis of some medicinal plants used in traditional medicine by Atomic Absorption Spectrophotometer (AAS). *Journal of Medicinal Plants Research* 4(19).