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THE MOLECULAR PERSPECTIVE OF MANIPURI RATS

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ABSTRACT

Manipur a tiny state of India is a part of the Indo-Burma hotspots of the World and diversity may be due to the connectivity through the mountain range with the South Eastern Asia particularly the rat species. The diversity of rats of the Manipur remains largely unexplored besides some of the works of the Zoological Survey of India. Here we present the two rat species that were ascertain through the COI (719 bp) sequences, namely *Rattus nitidus* and *Rattus tanezumi*. The diploid of these two species is 42 as reported and grouped as *Rattus tanezumi* group. How much difference from the rest of the world is yet to be ascertained?

Keywords: Manipur, First Record, *Rattus Nitidus*, *Rattustanezumi*, Oriental Rat, Heterochromatic Q-Arms

Introduction

The rats of Manipur have evolutionary importance as this land is in the junction of Indo-Burma hotspots and South East Asia that overlaps with four hotspots of biodiversity containing exceptional concentrations of endemic species and facing huge habitat loss (Myers *et al.*, 2000). The representatives of the genus *Rattus* have been confirmed to be arise from the South Asian countries and Indian continent is regarded as old habitat of the genus. Some of the taxonomic studies of the rats in India had been initiated through the leadership of Zoological Survey of India (ZSI), Kolkata. The diversity of rats of the Manipur, the easternmost state of India remains largely unexplored besides some of the works of the Zoological Survey of India. Out of a total of 4,629 species of mammals known in the world, 372 species occur in the Indian union. Of these, 69 species are reported from Manipur (Mandal *et al.*, 2005). Five subspecies under the genus *Rattus* viz., *Rattus rattus brunneusculus* (Hodgson), *Rattus rattus bullocki* Roonwal, *Rattus rattus tistae* Hinton, *Rattus nitidus obsoletus* Hinton, and *Rattus manipulus manipulus* (Thomas) besides one species *Rattus mackenziei* (Thomas) had been reported from Manipur. *Rattus mackenziei* (Thomas) and *Rattus manipulus manipulus* (Thomas) were included in the genus *Berylmys* as *Berylmys mackenziei* (Thomas) and *Berylmys manipulus* (Thomas) respectively. In this paper we are reporting the occurrence of two more species of the genus *Rattus* viz. *Rattus tanezumi* (Temminck, 1844) and *Rattus nitidus*, Hinton for the first time from Manipur.

MATERIALS AND METHODS

18 (12 males and 6 females) rat specimens of two different species of the genus *Rattus* were procured death by either killed using slang or poison or alive by trapping and they were taken to the laboratory. Each specimen was given a code of PBG with numbers and the skull and skin were deposited in the University Museum after the experiments. The morphological studies were carried out according to Agrawal (2000), Aplin *et al.*, (2003) and Alfred (2005). The skull preparation was done according to Herbreteau *et al.*, (2011); chromosomes were harvested from the bone marrow cells of femur using standard colchicine-hypotonic-air drying technique. Approval of the Institutional Ethics Committee (IEC), Central Agricultural University was obtained for using live animals and protocols of the IEC were followed throughout the study. The chromosomes were stained with Giemsa for conventional karyotyping. Either liver or kidney tissues were used for DNA extraction by phenol-chloroform technique.

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Taxonomic Backgrounds

Morphological Characters of the *Rattus Nitidus*

In this species the fur is soft and quite thick, particularly in the typical race. The tail is nearly naked, that with signs of growth of hairs throughout its length. Hind foot with five toes, all clawed; the hallux shorter than the fifth, which is shorter than the central three; apparently six plantar pads. Tail little longer than head and body as rule (107% average for typical race, Hinton; 99% average for *obsoletus*). Mammary 12, as rule. The back with is usually dark brown, occasionally with a darker mid-dorsal patch or line. Feet usually yellowish or whitish, rarely dark. Tail normally wholly dark (Ellerman, 1961). *Rattus nitidus* had Harsher-furred; bullae apparently under 15% of the occipitonasal length; tail averages just shorter than head and body. Occipitonasal length not exceeding 45.5 mm. Diastema proportionately shorter than in *Rattus bowersi*.

Morphological Characters of the *Rattus Tanezumi*

All the measurements were taken from nine fully grown rats collected from Karang, Loktak Lake. Medium-sized rats, dorsal fur slightly reddish grey brown. Both pes and manus had a darker colouration unto the toes. Ventral part of the rat was purely white with clear demarcation between dorsal and ventral that extended onto the cheek with an intervention of smoky T-shaped around throat and thoracic region as on the upper parts of manus and pes in colouration.

Cytological Characters of *Rattus Nitidus* and *Rattus Tanezumi*

The 2n of *Rattus nitidus* is 42. The karyotype comprises one pair of subtelocentric (#1), 12 pairs of acrocentrics pairs (#2-13), seven pairs of small metacentric pairs (#14-20) as autosomal compliments. The sex chromosomes are subtelocentrics (Figure 1 B). The q-arms of the X chromosomes were heterochromatic. All the 15 rats available for karyotyping showed without exception 42 chromosomes in which chromosome numbers 14-20 were small metacentric chromosomes and other autosomal and sex chromosomes were acrocentrics. The X chromosome was 10th or 11th of autosomal chromosome in length and Y chromosome was the shortest in the genome (Figure 1A).

Molecular Identity of *Rattus Nitidus* and *Rattus Tanezumi*

For confirmation of the molecular identity of the two rats the barcoding gene, cytochrome oxidase subunit I (COI) was sequenced with the primers viz., BatL5310 (5' ACTTCTGGGTGTCCAAAGAATCA 3') and R6036R (5' CCTACTCRGCCATTTTACCTATG 3'). PCR amplification was carried out in 35µl reaction volume with 50 ng of DNA, 0.2 µM of primer pairs BatL5310 and R6036R, 200 µM of dNTP mix, 1.5 mM MgCl₂ and 1 Unit of Taq DNA polymerase (Qiagen). The thermal cycling parameters were 4 min at 94°C, 40 cycles of 30 s at 94°C, 30 s at 48°C, 60 s at 72°C, and a final extension of 10 min at 72°C. The PCR products were sequenced bi-directionally and the consensus sequences were compared with homologous sequences available with www.ceropath.org/trees developed may be see-Supplementary I). The two sequences of *Rattus nitidus* and *R. tanezumi* were deposited to the NCBI GenBank with accession numbers JQ906931 and JQ918374. For phylogenetic analysis and cladistic grouping, sequences from NCBI for related species were taken and a phylogenetic tree drawn using MEGA 5.0. Eleven more COI sequences in the Fasta format were downloaded from the NCBI website and the phylogenetic tree to compare the status of the two taxa (Supplementary II).

RESULTS

The evolutionary history was inferred by using the Maximum Likelihood method based on the Kimura 2-parameter model [1]. The tree with the highest log likelihood (-1546.4237) is shown. The percentage of trees in which the associated taxa clustered together is shown next to the branches. Initial tree(s) for the heuristic search were obtained automatically by applying Neighbor-Join and BioNJ algorithms to a matrix of pairwise distances estimated using the Maximum Composite Likelihood (MCL) approach, and then selecting the topology with superior log likelihood value. A discrete Gamma distribution was used to model evolutionary rate differences among sites (5 categories (+G, parameter = 0.3831)). The tree is drawn to scale, with branch lengths measured in the number of substitutions per site. The analysis involved 12 nucleotide sequences. Codon positions included were 1st+2nd+3rd+Noncoding. All positions

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with less than 50% site coverage were eliminated. That is, fewer than 50% alignment gaps, missing data, and ambiguous bases were allowed at any position. There were a total of 675 positions in the final dataset. Evolutionary analyses were conducted in MEGA5 [2].

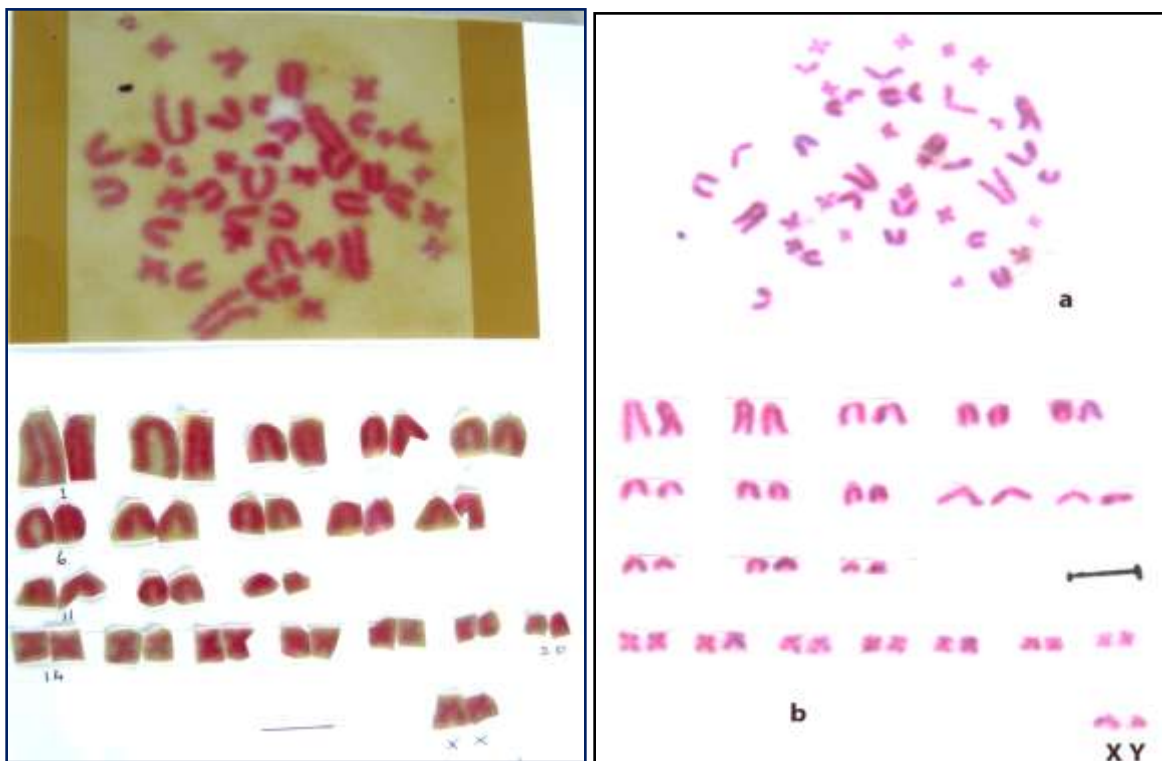


Figure 1: (a) Metaphase and karyotype of *Rattus tanezumi*; (b) Metaphase and karyotype of *Rattus nitidus*

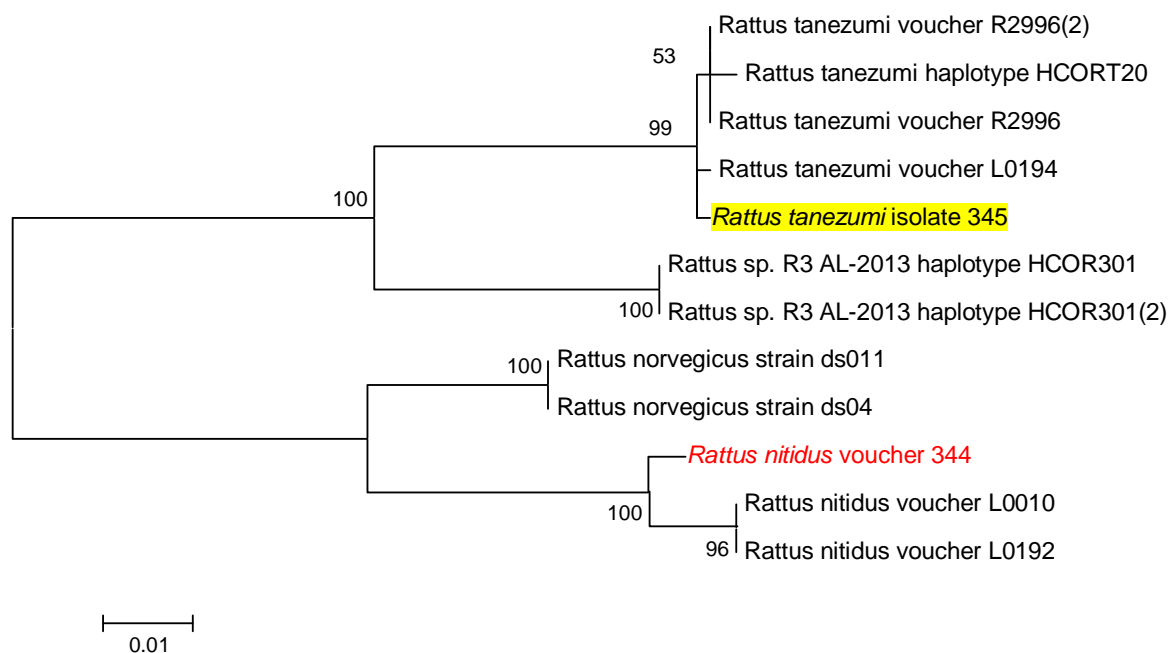


Figure 2: Molecular Phylogenetic analysis by Maximum Likelihood method

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The above clearly shows the validity of the species of the Manipur as distinct taxon as compared to different parts of the world but two sequences cannot be taken as the representative of the various species of Manipur. So much work is needed for explaining.

DISCUSSION

Rattus nitidus, also called the Himalayan rat, is indigenous to mainland Southeast Asia and occurs in south China (including Hainan Island), Vietnam, Laos, northern Thailand, Burma, Bangladesh, Nepal, Bhutan, and northern India; it is also found on the islands of central Sulawesi, Luzon island in the Philippines, Pulau Seram in the Mollucas, the Vogelkop Peninsula of the Province of Papua, and the Palau Islands, probably due to human-mediated introductions (Aplin *et al.*, 2003). Li *et al.*, (2008) reported the $2n=42$ of *Rattus nitidus* comprising of eight metacentric pairs, two subtelocentric pairs, and 10 acrocentric pairs as autosomal complements and acrocentrics XX from China. In the present study also the karyotype is consisting of ten acrocentric pairs (#2-8, 10-12), two subtelocentric pairs (#1, and 9), eight pairs of small metacentric pairs (#13, 14-20) and acrocentrics X and Y. Hence the *Rattus nitidus* reported from Manipur is quite similar to the Chinese species even though the chromosome numbers are different. Differences in the karyotype may be due to adoption of different preparation methods or by interpretative differences among studies.

The basic chromosome number of the black rat *Rattus rattus* is 42 (21 pairs) consisting of 13 acrocentric pairs, 7 metacentric autosomal pairs and acrocentric X and Y; pairs 1, 9 and 13 are polymorphic due to acrocentric or subtelocentric (metacentric) members. Similarities in length and G-banding pattern between the acrocentric and subtelocentric, and also the frequencies and distribution patterns of the polymorphic chromosomes strongly suggest origin of the subtelocentrics by the pericentric inversion of the acrocentrics (Yosida, 1983). Subtelocentric pair 1, 9 and 13 of *R. nitidus* could have been derived by pericentric inversion of the original acrocentric pair. The change of subtelocentric X chromosome pairs might involve the duplication of the heterochromatins in the genome as reported in *Niviventer* (Wang *et al.*, 2003).

Hinton (1919) differentiated *R. nitidus obsoletus* from *Rattus nitidus nitidus* on the colour of the under surface of the body, being with grey with a rusty tinge in the former and silvery in the latter. Ellerman (1961) mentioned that this colour-difference seems to be not constant and provisionally differentiated the two subspecies on the length of tail, shorter tail in *obsoletus* (99% of HB) and longer in *nitidus* (107% of HB).

An appraisal of the specimens of *Rattus nitidus* present in ZSI reveals that there is no clear-cut difference between *R. nitidus* and *R. obsoletus* either in the colour of the under surface of the body or in the length of tail. Hence, the subspecies *R. obsoletus* is being treated as a synonym of *Rattus nitidus nitidus* (Agrawal, 2000).

According to the above points it is difficult to come to a conclusion but on the basis of karyotype it will be wise enough to differentiate the two species into as *Rattus nitidus nitidus* from Mao and *Rattus nitidus obsoletus* from the Ukhrul. The soft palate ridges strengthened the differentiation of the two species. One thing is certain from the observation that presence of subtelocentric chromosomes there is tendency of having the much darker fur hairs or melanistic forms. Future work should be with more numbers of the specimens and more study sites.

Rattus tanezumi is the oldest name for the $2n = 42$ group of Asian house rats that is distinguished from the $2n = 38/40$ *Rattus rattus* not only by chromosomal characters but also by morphological and biochemical traits (Wilson and Reeder, 2005). In Manipur the rat is not common in valley but mostly concentrated on the hillock of Karang island in the Loktak Lake. This species is morphologically quite similar to *Rattus rattus* and is very difficult to identify. Hence a barcoding gene (mitochondrial COI region) was sequenced for identification purpose. The species is listed as Least Concern due to its wide distribution and tolerance of a wide variety of habitat types (Heaney and Molur, 2008).

Yosida and Sagai (1973) reported subtelocentric chromosome number 1 under the name *Rattus rattus tanezumi* (now named *R. tanezumi*; Wilson and Reeder, 2005) but in our studies (Dhananjoy *et al.*, 2012)

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the chromosome is acrocentric. They also suggested that subtelocentric chromosomes were derived from acrocentric chromosomes through pericentric inversion. The chromosome numbers 1, 9 and 13 are the hot spots for chromosome polymorphism in genus *Rattus*, but none of these chromosomes are either heteromorphic or homomorphic due to subtelocentric and acrocentric. Instead we found chromosome numbers 15, 19 and 20 to be heteromorphic due to metacentric and submetacentrics. According to Yosida and Sagai (1972), some *Rattus* species having few small metacentrics are included in the third group which might be ancestral type of other species with 7 metacentrics. From the above discussion it can be concluded that *Rattus tanezumi* is ancestral to *Rattus rattus* species in order of subtelocentric and small metacentric chromosomes of autosomes and further intensive works should be carried out in order to decide the facts.

Wilson and Reeder (2005) synonymised *Rattus brunneusculus* (Hogson, 1945) with *Rattus tanezumi*. In Indian context also *Rattus brunneusculus* was reported from Manipur instead of the *Rattus tanezumi* (Alfred, 2005) and Ellerman (1961) from The Loktak Lake. Alfred reports that the *brunneusculus* had longer tail, being 123-131% of head and body length but in our study it is 90%. Again Ellerman (1961) synonymised it with *sikkimensis* and argued that the typical *brunneusculus* has the underparts yellowish, not very typical for a wild race, besides this the characteristic feature of both *brunneusculus* and *sikkimensis* is “the bullae are usually less than 17% of the occipitonasal length” but in our study it is 19.51%. So the true taxonomic position of *Rattus tanezumi* is yet to be decided particularly in Indian context. A further study involving the three species through morphology, cytology and molecular approach is suggested to resolve the taxonomic conflicts of these three species in future.

The present trend in the taxonomy of *Rattus* is rather changing. According to Aplin *et al.*, (2011) *Rattus rattus* (Lineage I) has the diploid number of 38 whereas *Rattus tanezumi* has $2n=42/40$. They grouped the entire Black Rats into *Rattus rattus* and *Rattus tanezumi* based on the DNA sequences for taxonomic rationalization. So the two rats from Manipur belong to the *R. tanezumi* group. According to them India is the long term resident with stable population of Lineage I and the western Indochina and uplands of eastern Indochina is regarded as the likely geographic focus of diversification of Lineage II. The NE region of India is a part of the Indochina hence this region is a precious site both for invention of new species and determine the focus of diversification of rats. The future works on the taxonomic supported by molecular evidences will yield exciting results from this tiny area of the India.

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Supplimentary I



DNA Sequence Identification Results _ CERoPath - Barcoding Rodent South East Asia 5.pdf

Supplimentary II

Sequences of the 13 Specimens used in the study

>1. *Rattus nitidus* voucher L0010 (COXI) mitochondrial

CCCTCTACCTATTATTTGGAGCCTGAGCAGGAATAGTAGGAACAGCTTTAAGTATTCTAATT
CGAGCTGA
ATTAGGACAACCAGGCGCACTCCTAGGCGATGACCAAATCTATAATGTCATCGTCACAGCCC
ATGCATTC
GTAATAATCTTCTTTATAGTAATACCTATAATAATCGGAGGCTTCGGAAACTGACTTGTACC
ACTAATAA
TTGGAGCCCCCTGATATGGCATTCCCACGAATAAATAACATGAGTTTTTGTACTGCTCCCTCCAT
CATTCT
ACTTCTTTTAGCATCCTCCATAGTAGAAGCCGGAGCTGGGACAGGATGGACTGTATATCCTC
CCTTAGCC
GGAAACCTAGCCCATGCTGGAGCATCCGTAGACTTAACTATTTTTTCCCTCCACCTAGCTGGT
GTGTCTT
CTATCTTAGGAGCTATCAACTTTATTACCACTATCATTAACATGAAACCCCCTGCTATAACCC
AGTATCA
GACACCTCTCTTTGTATGATCCGTGCTAATTACAGCTGTCCTACTACTTCTCTCCCTACCCGT
GTTAGCA
GCAGGCATCACTATGCTCCTTACAGACCGAAATTTAAATACTACCTTCTTCGATCCCGCTGG
AGGGGGAG
ACCCAATTCTTTATCAACACCTATT

>2. *Rattus tanezumi* voucher R4481 (COXI) mitochondrial

CCCTCTATCTATTATTTGGTGCCTGAGCAGGAATAGTAGGAACAGCCTTGAGCATTCTAATTC
GAGCTGA
ACTAGGACAACCAGGAGCACTCCTAGGCGATGACCAAATTTATAATGTCATTGTTACAGCCC
ATGCATTC
GTAATAATTTTCTTTATAGTTATGCCTATGATAATCGGAGGCTTCGGAAACTGGCTTGTACCA
CTAATGA
TTGGAGCCCCCTGATATAGCATTCCCACGAATAAACAATATAAGCTTTTGATTGCTTCCCCCAT
CATTTTT
ACTCCTTTTAGCATCATCTATAGTAGAAGCCGGAGCCGGAACAGGATGAACAGTATACCCGC
CCTTAGCC
GGTAACCTAGCCCATGCCGGAGCATCCGTTGACCTAACCATTTTCTCCCTTCACCTAGCTGGT
GTATCCT
CTATCTTAGGAGCTATTAATTTTATCACCCTATCATCAATATAAAACCCCCTGCTATAACCC
AATATCA
GACCCCTCTATTTGTGTGATCCGTATTAATTACAGCTGTACTTCTACTTCTTCACTACCAGTT
TTAGCA
GCAGGCATTACCATACTCCTCACAGATCGAAACCTAAATACTACTTTTTTTGATCCTGCTGGA
GGCGGAG
ATCCAATTCTCTATCAACATCTATT

>3. *Rattus tanezumi* voucher R3122 (COXI) mitochondrial

AGGAACAGCCTTGAGCATTCTAATTCGAGCTGAACTAGGACAACCAGGAGCACTCCTAGGC
GATGACCAA

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ATTTATAATGTCATTGTTACAGCCCATGCATTTCGTAATAATTTTCTTTATAGTTATGCCTATG
ATAATCG
GAGGCTTCGGAAACTGGCTTGTACCACTAATGATTGGAGCCCCTGATATAGCATTCCCACGA
ATAAACAA
TATAAGCTTTTGATTGCTTCCCCCATCATTTTTACTCCTTTTAGCATCATCTATAGTAGAAGCC
GGAGCC
GGAACAGGATGAACAGTATACCCACCCTTAGCCGGTAACCTAGCCCATGCCGGAGCATCCG
TTGACCTGA
CCATTTTCTCCCTTCACCTAGCTGGTGTATCCTCTATCTTAGGAGCTATTAATTTTATCACCAC
TATCAT
CAACATAAAACCCCTGCTATAACCCAATATCARACCCCTCTATTTGTGTGATCCGTATTAAT
TACAGCT
GTACTTCTACTTCTTTCACTACCAGTTTTAGCAGCAGGCATTACCATACTCCTCACAGATCGA
AACCTAA
ATACTACTTTTTTTTGATCCTGCTGG

>4. *Rattus tanezumi* voucher R2996 (COXI) mitochondrial

CCCTCTATCTATTATTTGGTGCCTGAGCAGGAATAGTAGGAACAGCCTTGAGCATTCTAATTC
GAGCTGA
ACTAGGACAACCAGGAGCACTCCTAGGCGATGACCAAATTTATAATGTCATTGTTACAGCCC
ATGCATTC
GTAATAATTTTCTTTATAGTTATGCCTATGATAATCGGAGGCTTCGGAAACTGGCTTGTACCA
CTAATGA
TTGGAGCCCCTGATATAGCATTCCCACGAATAAACAATATAAGCTTTTGATTGCTTCCCCCAT
CATTTTT
ACTCCTTTTAGCATCATCTATAGTAGAAGCCGGAGCCGGAACAGGATGAACAGTATACCCAC
CCTTAGCC
GGTAACCTAGCCCATGCCGGAGCATCCGTTGACCTAACCATTTTCTCCCTCCACCTAGCTGGT
GTATCCT
CTATCTTAGGAGCTATTAATTTTATCACCCTATCATCAATATAAAACCCCTGCTATAACCC
AATATCA
GACCCCTCTATTTGTGTGATCCGTATTAATTACAGCTGTACTTCTACTTCTTTCACTACCAGTT
TTAGCA
GCAGGCATTACCATACTCCTCACAGATCGAAACCTAAATACTACTTTTTTTTGATCCTGCTGGA
GGCGGAG
ATCCAATTCTCTATCAACATCTATT

>5. *Rattus tanezumi* voucher R0130 (COXI) mitochondrial

CCCTCTATCTATTATTTGGTGCCTGAGCAGGAATAGTAGGAACAGCCTTGAGCATTCTAATTC
GAGCTGA
ACTAGGACAACCAGGAGCACTCCTAGGCGATGACCAAATTTATAATGTCATTGTTACAGCCC
ATGCATTC
GTAATAATTTTCTTTATAGTTATGCCTATGATAATCGGAGGCTTCGGAAACTGGCTTGTACCA
CTAATGA
TTGGAGCCCCTGATATAGCATTCCCACGAATAAACAATATAAGCTTTTGATTGCTTCCCCCAT
CATTTTT
ACTCCTTTTAGCATCATCTATAGTAGAAGCCGGAGCCGGAACAGGATGAACAGTATACCCAC
CCTTAGCC
GGTAACCTAGCCCATGCCGGAGCATCCGTTGACCTAACCATTTTCTCCCTTCACCTAGCTGGT
GTATCCT
CTATCTTAGGAGCTATTAATTTTATCACCCTATCATCAATATAAAACCCCTGCTATAACCC
AATATCA

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GACCCCTCTATTTGTGTGATCCGTATTAATTACAGCTGTACTTCTACTTCTTTCACTACCAGTT
TTAGCA

GCAGGCATTACCATACTCCTCACAGATCGAAACCTAAATACTACTTTTTTTGATCCTGCTGGA
GGCGGAG

ATCCAATTCTCTATCAACATCTATT

>6. *Rattus rattus* voucher MDZ10 (COXI) mitochondrial

CCCTCTATCTATTATTTGGTGCCTGAGCAGGAATAGTAGGAACAGCCTTAAGTATTCTCATTC
GAGCTGA

ACTAGGACAACCAGGAGCACTCCTAGGCGATGACCAAATTTATAATGTCATTGTTACAGCCC
ATGCATTC

GTAATAATTTTCTTTATAGTTATGCCTATGATAATCGGAGGCTTCGGAAACTGGCTTGTGCCG
CTAATGA

TTGGAGCCCCCTGATATAGCATTCCCACGAATAAACAACATAAGCTTTTGATTACTTCCCCCAT
CATTTTT

ACTCCTTTTAGCATCATCTATGGTAGAAGCCGGAGCCGGAACAGGATGAACAGTATACCCAC
CCTTAGCC

GGTAATCTAGCCCATGCCGGAGCATCTGTTGATCTAACCATTTTCTCCCTTCACCTAGCCGGC
GTATCCT

CTATCTTAGGAGCTATTAATTTTATCACCCTATTATCAATATAAAACCCCCTGCTATAACCC
AATATCA

GACACCTCTATTTGTGTGATCCGTATTAATTACAGCTGTACTTCTACTTCTTTCACTACCAGT
GTTGGCA

GCAGGCATTACCATACTCCTCACAGATCGAAACCTAAATACTACTTTTTTTGATCCTGCTGGA
GGCGGAG

ATCCAATTCTCTATCAACATCTATT

>7. *Rattus nitidus* voucher L0192 (COXI) mitochondrial

CCCTCTACCTATTATTTGGAGCCTGAGCAGGAATAGTAGGAACAGCTTTAAGTATTCTAATT
CGAGCTGA

ATTAGGACAACCAGGCGCACTCCTAGGCGATGACCAAATCTATAATGTCATCGTCACAGCCC
ATGCATTC

GTAATAATCTTCTTTATAGTAATACCTATAATAATCGGAGGCTTCGGAAACTGACTTGTACC
ACTAATAA

TTGGAGCCCCCTGATATGGCATTCCCACGAATAAATAACATGAGTTTTTGACTGCTCCCTCCAT
CATTTCT

ACTTCTTTTAGCATCCTCCATAGTAGAAGCCGGAGCTGGGACAGGATGGACTGTATATCCTC
CCTTAGCC

GGAAACCTAGCCCATGCTGGAGCATCCGTAGACTTAACTATTTTTTCCCTCCACCTAGCTGGT
GTGTCTT

CTATCTTAGGAGCTATCAACTTTATTACCACTATCATTAAACATGAAACCCCCTGCTATAACCC
AGTATCA

GACACCTCTCTTTGTATGATCCGTGCTAATTACAGCTGTCCTACTACTTCTCTCCCTACCCGT
GTTAGCA

G

>8. *Rattus nitidus* voucher L0180 (COXI) mitochondrial

CCCTCTACCTATTATTTGGAGCCTGAGCAGGAATAGTAGGAACAGCTTTAAGTATTCTAATT
CGAGCTGA

ATTAGGACAACCAGGCGCACTCCTAGGCGATGACCAAATCTATAATGTCATCGTCACAGCCC
ATGCATTC

GTAATAATCTTCTTTATAGTAATACCTATAATAATCGGAGGCTTCGGAAACTGACTTGTACC
ACTAATAA

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TTGGAGCCCCTGATATGGCATTCCCACGAATAAATAACATGAGTTTTTGGCTGCTCCCTCCAT
CATTCT
ACTTCTTTTAGCATCCTCCATAGTAGAAGCCGGAGCTGGGACAGGATGGACTGTATATCCTC
CCTTAGCC
GGAAACCTAGCCCATGCTGGAGCATCCGTAGACTTAACTATTTTTTCCCTCCACCTAGCTGGT
GTGTCTT
CTATCTTAGGAGCTATCAACTTTATTACCACTATCATTAACATGAAACCCCCTGCTATAACCC
AGTATCA
GACACCTCTCTTTGTATGATCCGTACTAATTACAGCTGTCCTACTACTTCTCTCCCTACCCGT
GTTAGCA
GCAGGCATCACTATGCTCCTTACAGACCGAAATTTAAATACTACCTTCTTCGATCCCGCTGG
AGGGGGAG
ACCCAATTCTTTATCAACACCTAT

>9. *Rattus tanezumi* voucher L0100 (COXI) mitochondrial

CCCTCTATCTATTATTTGGTGCCTGAGCAGGAATAGTAGGGACAGCCTTGAGCATTCTAATTC
GAGCTGA
ACTAGGACAACCAGGAGCACTCCTAGGCGATGACCAAATTTATAATGTCATTGTTACAGCCC
ATGCATTC
GTAATAATTTTCTTTATAGTTATGCCTATGATAATCGGAGGCTTCGGAAACTGGCTTGTACCA
CTAATGA
TTGGAGCCCCTGATATAGCATTCCCACGAATAAACAATATAAGCTTTTGATTGCTTCCCCCAT
CATTTTT
ACTCCTTTTAGCATCATCTATAGTAGAAGCCGGAGCCGGAACAGGATGAACAGTATACCCGC
CCTTAGCC
GGTAACCTAGCCCATGCCGGAGCATCCGTTGACCTAACCATTTTCTCCCTTCACCTAGCTGGT
GTATCCT
CTATCTTAGGAGCTATTAATTTTATCACCCTATCATCAATATAAAACCCCCTGCTATAACCC
AATATCA
GACCCCTCTATTTGTGTGATCCGTATTAATTACAGCTGTACTTCTACTTCTTTCACTACCAGTT
TTAGCA
GCAGGCATTACCATACTCCTCACAGATCGAAACCTAAATACTACTTTTTTTGATCCTGCTGGA
GGCGGAG
ATCCAATTCTCTATCAACATCTAT

>10. 344 *Rattus nitidus*

ACCTATGTTTCGTAAACCGTTGACTCTTTTCAACTAACCACAAAGAT
ATCGGAACCCCTCTACCTATTATTTGGAGCCTGAGCAGGAATAGTAGGAACAGCTTTAAGTAT
TCTAATTC
GAGCTGAATTAGGTCAACCAGGCGCACTCCTAGGCGATGACCAAATCTATAATGTCATCGTC
ACAGCCCA
TGCATTTCGTAATAATTTTCTTTATAGTAATACCTATAATAATCGGAGGCTTCGGAAACTGACT
TGTGCCA
CTAATAATTGGAGCCCCTGATATAGCATTCCCACGAATAAATAACATGAGTTTTTGGCTGCT
CCCTCCAT
CATTTCTACTTCTTTTAGCATCCTCCATAGTAGAAGCCGGAGCTGGAACAGGATGGACTGTA
TATCCTCC
CTTAGCCGGAAACCTAGCCCATGCTGGAGCATCCGTAGACTTAACTATTTTTTCCCTCCACCT
AGCTGGT
GTGTCTTCTATCTTAGGAGCTATCAACTTTATTACCACTATCATTAACATGAAACCCCCTGCT
ATAACCC

Research Article

AGTATCAGACACCTCTCTTTGTATGATCCGTGCTAATTACAGCTGTCCTACTACTTCTCTCCC
TACCCGT
GTTAGCAGCAGGTATCACCATACTCCTTACAGACCGAAATTTAAATACTACCTTCTTCGACC
CCGCTGGA
GGGGGAGACCCAATTCTTTATCAACACCTATTCTGATTCTTTG

>11. 345 *Rattus tanezumi*

ACCTATGTTTCGTAAACCGTTGACTCTTTTCAACTAACCACAAAGATATCGGAACCCTCTATCT
ATTATTT
GGTGCCTGAGCAGGAATAGTAGGAACAGCCTTGAGCATTCTAATTCGAGCTGAACTAGGAC
AACCAGGAG
CACTCCTAGGCGATGACCAAATTTATAATGTCATTGTTACAGCCCATGCATTTCGTAATAATTT
TCTTTAT
AGTTATGCCTATGATAATCGGAGGCTTCGGAACTGGCTTGTACCACTAATGATTGGAGCCC
CTGATATA
GCATTCCCACGAATAAACAATATAAGCTTTTGATTGCTTCCCCCATCATTTTTACTCCTTTTA
GCATCAT
CTATAGTAGAAGCCGGAGCCGGAACAGGATGAACAGTATACCCACCCTTAGCCGGTAACCT
AGCCCATGC
CGGAGCATCCGTTGACCTAACCATTTTCTCCCTTCACCTAGCTGGTGTATCCTCTATCTTAGG
AGCTATT
AATTTTATCACCCTATCATCAATATAAAACCCCTGCTATAACCCAATATCAGACCCCTCTA
TTTGTGT
GATCCGTATTAATTACAGCCGTA CTCTACTTCTTTCACTACCAGTTTTAGCAGCAGGCATTA
CCATACT
CCTCACAGATCGAAACCTAAATACTACTTTTTTTGATCCTGCTGGAGGCGGAGATCCAATTCT
CTATCAA
CATCTATTTTGATTCTTTG

>12. *Rattus norvegicus isolate 2 cytochrome oxidase subunit mitochondrial*

AACCCTCTACCTATTATTTGGAGCCTGAGCAGGAATAGTAGGGACAGCTTTAAGTATTCTAA
TTCGAGCT
GAACTAGGACAGCCAGGCGCACTCCTAGGAGATGACCAAATCTATAATGTCATCGTCACAG
CCCATGCAT
TCGTAATAATTTTCTTTATAGTAATACCTATAATAATTGGAGGCTTCGGGAAGTACTTGTAC
CACTAAT
AATTGGAGCCCCTGATATAGCATTCCACGAATAAATAACATAAGCTTTTGACTGCTTCCTC
CATCATTT
CTACTCCTTTTAGCATCCTCCATAGTAGAAGCTGGAGCTGGAACAGGATGAACAGTATATCC
CCCCTTAG
CCGGAACCTAGCCCATGCTGGAGCATCCGTAGATTTAACTATTTTTTCCCTCCACCTAGCCG
GGGTGTC
TTCTATCTTAGGAGCTATCAACTTTATCACCCTATCATTAATATAAAACCCCTGCTATAAC
CCAATAT
CAGACACCTCTCTTTGTATGATCCGTACTAATTACAGCCGTCCTACTACTTCTCTCACTGCCA
GTATTAG
CAGCAGGTATCACTATACTCCTTACAGACCGAAATCTAAATACTACTTTCTTCGACCCCGCTG
GAGGTGG
AGACCAATCCTTTATCAACACCTATTC

>13. *Rattus norvegicus mitochondrial*

COITTCATAAACCGTTGACTCTTTTCAACT
AACCACAAAGATATCGGAACCCTCTACCTATTATTTGGGGCCT

Research Article

GAGCAGGAATAGTAGGGACAGCTTTAAGTATTCTAATTCGAGCTGAACTAGGGCAGCCAGG
TGCACTCCT
AGGAGATGACCAAATCTATAATGTCATCGTCACAGCCCATGCATTCGTAATAATTTTCTTTAT
AGTAATA
CCTATAATAATTGGAGGCTTCGGAAACTGACTTGTCCCACTAATAATTGGAGCCCCTGATAT
AGCATTCC
CACGAATAAATAACATAAGCTTTTGACTGCTTCCTCCATCGTTTCTACTCCTTTTAGCATCCT
CCATAGT
AGAAGCTGGAGCTGGAACAGGATGAACAGTATATCCCCCCTTAGCCGGAAACCTAGCCCAT
GCTGGAGCA
TCCGTAGATTTA ACTATTTTTTCCCTCCACCTAGCCGGGGTGTCTTCTATCTTAGGAGCTATC
AACTTTA
TCACCACTATCATTAATATAAAACCCCTGCTATAACCCAATATCAAACACCTCTCTTTGTAT
GATCCGT
ACTAATTACAGCCGTCCTACTACTTCTCTCACTGCCAGTATTAGCAGCAGGTATCACTATACT
CCTTACA
GACCGAAATCTAAATACTACTTTCTTCGACCCCGCTGGAGGTGGAGACCCAATTCTTTATCA
ACACCTAT
TC